

UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

REGION 4
ATLANTA FEDERAL CENTER
61 FORSYTH STREET
ATLANTA, GEORGIA 30303-8960

MAR 3 1 2014

CERTIFIED MAIL 7009 0960 0000 6489 3351 RETURN RECEIPT REQUESTED

Mr. Scott Stiles
Director
Public Works Department
City of Cornersville
118 South Main Street
Cornersville, Tennessee 37047

Re: Notice of Violation No. 309-2014-04 Information Request pursuant to Section 308 of the Clean Water Act National Pollutant Discharge Elimination System Permit No.: TN0061841 Diagnostic Evaluation Report

Dear Mr. Stiles:

The Diagnostic Evaluation Inspection report (Report), written for the City of Cornersville Wastewater Treatment Plant (City) inspection that was conducted by the U.S. Environmental Protection Agency Region 4's Science and Ecosystem Support Division on May 6-7, 2013, is included as an enclosure to this letter. The purpose of the inspection was to determine why effluent limit exceedances have occurred at the plant and to determine the City's compliance with the permit. The Report outlines several findings and also includes the following deficiencies that the City must address to ensure full compliance with its National Pollution Discharge Elimination System (Permit) Permit No.: TN0061841:

- 1. The 24 Hour composite samples were collected over a shorter time period than the 24 hours specified in the Permit (Part 1 of Permit);
- 2. The flow meter at outfall 001 was recording 25 percent less than the actual flow exceeding the requirement of within ± 10 percent of the primary flow measuring device (Part 1.2.1 of Permit);
- 3. The plant was experiencing the following operation and maintenance issues at the plant that affected compliance with the permit (Part 2.1.4 of Permit):
 - o The 1.2 million gallon equalization tank was near to exceeding capacity;
 - o The Ultra Violet disinfection lamps did not appear to be clean;
 - o The flow monitoring data reviewed within 15 months of the inspection showed that the plant was hydraulically overloaded for five months;
 - o There was not a preliminary treatment system (bar screen, grit removal) to remove solids and trash from the influent wastewater; and
 - Solids buildup was observed in effluent trough and Parshall flume crest which were affecting accuracy of flow measurement.

- 4. The plants sample collection and monitoring procedures were not representative of the monitored activity (Part 1.2.1. of the Permit);
 - o The pH buffer solution used for instrument calibration was expired;
 - o The influent composite sampler was located after the equalization tank which does not allow representative sampling of the influent; and
 - o The influent composite sampler tubing was clogged.

Pursuant to Section 309(a)(1) of the Clean Water Act (CWA), 33 U.S.C. §1319(a)(1), the EPA also hereby notifies the City that on numerous occasions the City violated its Permit as indicated by the effluent limit exceedances identified in Enclosure A.

The EPA requests, pursuant to Section 308 of the CWA, 33 U.S.C. § 1318, that the City provide a written response for the inspection deficiencies noted above as well as each of the effluent limit exceedances listed in Enclosure A. The City's response should include a written explanation of the reasons for each of the aforementioned violations and a summary of actions taken or planned by the City to correct the problems and to prevent future violations. In instances where the actions are planned, please include a schedule for completing the actions. The City's written response to the EPA shall be due within 30 days from receipt of this letter. The submittal must be addressed to:

Ms. Alenda Johnson
U.S. Environmental Protection Agency, Region 4
Clean Water Enforcement Branch
61 Forsyth Street, S.W.
Atlanta, Georgia 30303-8960

The City's response should specifically reference the particular element and page number of the Report and should be organized for the purpose of clarity. In addition, all information submitted must be accompanied by the following certification signed by a responsible City official:

"I certify under penalty of law that this document and all attachments were prepared under my direction or supervision in accordance with a system designed to assure that qualified personnel properly gather and evaluate the information submitted. Based on my inquiry of the person or persons who manage the system, or those persons directly responsible for gathering the information, the information submitted is, to the best of my knowledge and belief, true, accurate, and complete. I am aware that there are significant penalties for submitting false information, including the possibility of fine and imprisonment for knowing violations."

Failure to comply with this information request may result in enforcement proceedings under Section 309 of the CWA, 33 U.S.C. § 1319, which could result in the judicial imposition of civil or criminal penalties or the administrative imposition of civil penalties. In addition, there is potential criminal liability for the falsification of any response to the requested information.

The City shall preserve, until further notice, all records (either written or electronic) that exist at the time of receipt of this letter that relate to any of the matters set forth in this letter. The term "records" shall be interpreted in the broadest sense to include information of every sort. The response to this information request shall include assurance that these record protection provisions were put in place, as required. No such records shall be disposed of until written authorization is received from the Chief of the Clean Water Enforcement Branch of the EPA, Region 4.

If you believe that any of the requested information constitutes confidential business information you may assert a confidentiality claim with respect to such information except for effluent data. Further details, including how to make a business confidentiality claim, are found in Enclosure C.

The State of Tennessee is being concurrently notified of these findings. The EPA is coordinating with the State to ensure that timely and appropriate enforcement action is taken and compliance with the conditions of the Permit is achieved.

If these findings are not resolved in a timely or appropriate manner, the EPA may take enforcement action, which may include issuance of an administrative order, assessment of administrative penalties, or initiation of a civil judicial action pursuant to Section 309 of the CWA, 33 U.S.C. § 1319.

If you have questions regarding this notice and information request, please contact Ms. Alenda Johnson, of my staff, at (404) 562-9761 or via e-mail at johnson.alenda@epa.gov.

Sincerely

James D. Giattina

Director

Water Protection Division

Enclosures

cc: Dr. Sandra Dudley, Director

Tennessee Department of Environment and Conservation

ENCLOSURE A

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Solids, total suspended	Monthly Avg. Min.	30.	22.2	mg/L	001G	01/31/2009
Solids, total suspended	Weekly Average	40.	44.2	mg/L	001G	01/31/2009
Solids, total suspended	Daily Maximum	45.	50.	mg/L	001G	01/31/2009
Solids, total suspended	Daily Maximum	37.5	59.5	lb/d	001G	01/31/2009
Solids, total suspended	Weekly Average	33.	120.5	lb/d	001G	01/31/2009
Total Ammonia Nitrogen	Monthly Average	1.9	13.06	mg/L	001G	01/31/2009
Total Ammonia Nitrogen	Weekly Average	2.4	19.43	mg/L	001G	01/31/2009
Total Ammonia Nitrogen	Daily Maximum	4.	21.11	mg/L	001G	01/31/2009
Total Ammonia Nitrogen	Monthly Average	1.6	11.36	lb/d	001G	01/31/2009
Total Ammonia Nitrogen	Daily Maximum	3.3	17.69	lb/d	001G	01/31/2009
Total Ammonia Nitrogen	Weekly Average	2.	10.9	lb/d	001G	01/31/2009
E. coli, MTEC-MF	Daily Maximum	941.	941.	#/100mL	001G	01/31/2009
BOD, carbonaceous, 05 day, 20 C	Monthly Average	10.	12.6	mg/L	001G	01/31/2009
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	20.	22.4	mg/Ľ	001G	01/31/2009
BOD, carbonaceous, 05 day, 20 C	Monthly Average	8.	10.8	lb/d	001G	01/31/2009
BOD, carbonaceous, 05 day, 20 C	Weekly Average	13.	14.4	lb/d	001G	01/31/2009
Solids, total suspended	Monthly Avg. Min.	30.	18.7	mg/L	001G	02/28/2009
Total Ammonia Nitrogen	Monthly Average	1.9	18.56	mg/L	001G	02/28/2009
Total Ammonia Nitrogen	Weekly Average	2.4	22.42	mg/L	001G	02/28/2009
Total Ammonia Nitrogen	Daily Maximum	4.	23.24	mg/L	001G	02/28/2009
Total Ammonia Nitrogen	Monthly Average	1.6	14.85	lb/d	001G	02/28/2009
Total Ammonia Nitrogen	Daily Maximum	3.3	28.2	lb/d	001G	02/28/2009
Total Ammonia Nitrogen	Weekly Average	2.	14.9	lb/d	001G	02/28/2009
E. coli, MTEC-MF	Monthly Geomean	126.	565.	#/100mL	001G	02/28/2009
E. coli, MTEC-MF	Daily Maximum	941.	5040.	#/100mL	001G	02/28/2009
BOD, carbonaceous, 05 day, 20 C	Monthly Average	10.	15.8	mg/L	001G	02/28/2009
BOD, carbonaceous, 05 day, 20 C	Weekly Average	15.	15.8	mg/L	001G	02/28/2009
BOD, carbonaceous, 05 day, 20 C	Monthly Average	8.	12.9	lb/d	001G	02/28/2009
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	16.7	24.58	lb/d	001G	02/28/2009
BOD, carbonaceous, 05 day, 20 C	Weekly Average	13.	15.2	lb/d	001G	02/28/2009
Solids, total suspended	Daily Maximum	45.	56.	mg/L	001G	03/31/2009
Solids, total suspended	Monthly Average	25.	25.3	,lb/d	001G	03/31/2009
Solids, total suspended	Daily Maximum	37.5	53.7	lb/d	001G	03/31/2009
Solids, total suspended	Weekly Average	33.	47.1	lb/d	001G	03/31/2009
Total Ammonia Nitrogen	Monthly Average	1.9	16.76	mg/L	001G	03/31/2009
Total Ammonia Nitrogen	Weekly Average	2.4	24.6	mg/L	001G	03/31/2009
Total Ammonia Nitrogen	Daily Maximum	4.	26.94	mg/L	001G	03/31/2009

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Total Ammonia Nitrogen	Monthly Average	1.6	12.68	lb/d	001G	03/31/2009
Total Ammonia Nitrogen	Daily Maximum	3.3	20.93	lb/d	001G	03/31/2009
Total Ammonia Nitrogen	Weekly Average	2.	12.7	lb/d	001G	03/31/2009
E. coli, MTEC-MF	Daily Maximum	941.	1100.	#/100mL	001G	03/31/2009
BOD, carbonaceous, 05 day, 20 C	Monthly Average	10.	17.3	mg/L	001G	03/31/2009
BOD, carbonaceous, 05 day, 20 C	Weekly Average	15.	17.3	mg/L	001G	03/31/2009
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	20.	23.4	mg/L	001G	03/31/2009
BOD, carbonaceous, 05 day, 20 C	Monthly Average	8.	13.5	lb/d	001G	03/31/2009
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	16.7	19.5	lb/d	001G	03/31/2009
BOD, carbonaceous, 05 day, 20 C	Weekly Average	13.	17.6	lb/d	001G	03/31/2009
Solids, total suspended	Weekly Average	40.	52.4	mg/L	001G	04/30/2009
Solids, total suspended	Daily Maximum	45.	380.	mg/L	001G	04/30/2009
Solids, total suspended	Monthly Average	25.	38.7	lb/d	001G	04/30/2009
Solids, total suspended	Daily Maximum	37.5	278.9	lb/d	001G	04/30/2009
Solids, total suspended	Weekly Average	33.	101.2	lb/d	001G	04/30/2009
Solids, settleable	Daily Maximum	1.	22.	, mL/L	001G	04/30/2009
Total Ammonia Nitrogen	Monthly Average	1.9	10.46	mg/L	001G	04/30/2009
Total Ammonia Nitrogen	Weekly Average	2.4	12.49	mg/L	001G	04/30/2009
Total Ammonia Nitrogen	Daily Maximum	4.	17.08	mg/L	001G	04/30/2009
Total Ammonia Nitrogen	Monthly Average	1.6	7.76	lb/d	001G	04/30/2009
Total Ammonia Nitrogen	Daily Maximum	3.3	11.97	lb/d	001G	04/30/2009
Total Ammonia Nitrogen	Weekly Average	2.	7.3	lb/d	001G	04/30/2009
Solids, total suspended	Monthly Avg. Min.	30.	28.9	mg/L	001G	05/31/2009
Solids, total suspended	Daily Maximum	37.5	38.3	lb/d	001G	05/31/2009
Solids, settleable	Daily Maximum	1.	3.5	mL/L	001G	05/31/2009
Total Ammonia Nitrogen	Monthly Average	1.1	7.95	mg/L	001G	05/31/2009
Total Ammonia Nitrogen	Weekly Average	2.	9.04	mg/L	001G	05/31/2009
Total Ammonia Nitrogen	Daily Maximum	2.5	15.57	mg/L	001G	05/31/2009
Total Ammonia Nitrogen	Monthly Average	.9	6.83	lb/d	001G	05/31/2009
Total Ammonia Nitrogen	Daily Maximum	2.1	7.07	lb/d	001G	05/31/2009
Total Ammonia Nitrogen	Weekly Average	1.7	7.07	lb/d	001G	05/31/2009
E. coli, MTEC-MF	Daily Maximum	941.	5400.	#/100mL	001G	05/31/2009
Solids, total suspended	Monthly Avg. Min.	30.	17.3	mg/L	001G	06/30/2009
Solids, total suspended	Monthly Avg. Min.	30.	9.	mg/L	001G	07/31/2009
Solids, total suspended	Monthly Avg. Min.	30.	7.3	mg/L	001G	08/31/2009
Total Ammonia Nitrogen	Monthly Average	1.1	2.86	mg/L	001G	08/31/2009
Total Ammonia Nitrogen	Weekly Average	2.	2.3	mg/L	001G	08/31/2009
Total Ammonia Nitrogen	Daily Maximum	2.5	11.09	mg/L	001G	08/31/2009
Total Ammonia Nitrogen	Monthly Average	.9	1.42	lb/d	001G	08/31/2009
BOD, carbonaceous, 05 day,						
20 C	Daily Maximum	20.	31.	mg/L	001G	08/31/2009

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Solids, total suspended	Monthly Avg. Min.	30.	11.4	mg/L	001G	09/30/2009
Total Ammonia Nitrogen	Monthly Average	1.1	1.61	mg/L	001G	09/30/2009
Total Ammonia Nitrogen	Daily Maximum	2.5	9.91	mg/L	001G	09/30/2009
Total Ammonia Nitrogen	Monthly Average	9	2.54	lb/d	001G	09/30/2009
Total Ammonia Nitrogen	Daily Maximum	2.1	2.52	lb/d	001G	09/30/2009
Total Ammonia Nitrogen	Weekly Average	1.7	2.52	lb/d	001G	09/30/2009
Solids, total suspended	Monthly Avg. Min.	30.	7.6	mg/L	001G	10/31/2009
Total Ammonia Nitrogen	Monthly Average	1.1	2.89	mg/L	001G	10/31/2009
Total Ammonia Nitrogen	Weekly Average	2.	2.83	mg/L	001G	10/31/2009
Total Ammonia Nitrogen	Daily Maximum	2.5	13.33	mg/L	001G	10/31/2009
Total Ammonia Nitrogen	Monthly Average	.9	3.61	lb/d	001G	10/31/2009
Total Ammonia Nitrogen	Daily Maximum	2.1	3.55	lb/d	001G	10/31/2009
Total Ammonia Nitrogen	Weekly Average	1.7	3.55	lb/d	001G	10/31/2009
E. coli, MTEC-MF	Daily Maximum	941.	2419.	#/100mL	001G	10/31/2009
Solids, total suspended	Monthly Avg. Min.	30.	13.3	mg/L	001G	11/30/2009
Solids, total suspended	Monthly Avg. Min.	30.	18.1	mg/L	001G	12/31/2009
Solids, total suspended	Daily Maximum	45.	52.5	mg/L	001G	12/31/2009
Solids, total suspended	Daily Maximum	37.5	67.9	lb/d	001G	12/31/2009
Solids, total suspended	Weekly Average	33.	54.9	lb/d	001G	12/31/2009
Solids, total suspended	Monthly Avg. Min.	30.	24.7	mg/L	001G	01/31/2010
Solids, total suspended	Daily Maximum	45.	56.	mg/L	001G	01/31/2010
Solids, total suspended	Daily Maximum	37.5	62.8	lb/d	001G	01/31/2010
Solids, total suspended	Weekly Average	33.	52.3	lb/d	001G	01/31/2010
Solids, settleable	Daily Maximum	1.	10.	mL/L	001G	01/31/2010
Total Ammonia Nitrogen	Monthly Average	1.9	3.38	mg/L	001G	01/31/2010
Total Ammonia Nitrogen	Weekly Average	2.4	7.28	mg/L	001G	01/31/2010
Total Ammonia Nitrogen	Daily Maximum	4.	8.34	mg/L	001G	01/31/2010
Total Ammonia Nitrogen	Monthly Average	1.6	2.86	lb/d	001G	01/31/2010
Total Ammonia Nitrogen	Daily Maximum	3.3	7.13	lb/d	001G	01/31/2010
Total Ammonia Nitrogen	Weekly Average	2.	2.5	lb/d	001G	01/31/2010
E. coli, MTEC-MF	Daily Maximum	941.	72914.	#/100mL	001G	01/31/2010
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	20.	20.9	mg/L	001G	01/31/2010
BOD, carbonaceous, 05 day, 20 C	Monthly Average	8.	8.1	lb/d	001G	01/31/2010
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	16.7	19.9	lb/d	001G	01/31/2010
BOD, carbonaceous, 05 day, 20 C	Weekly Average	13.	16.3	lb/d	001G	01/31/2010
Solids, suspended percent removal	Monthly Avg. Min.	60.	57.9	%	001G	01/31/2010
Solids, total suspended	Monthly Avg. Min.	30.	15.8	mg/L	001G	02/28/2010
Solids, total suspended	Daily Maximum	45.	51.5	mg/L	001G	02/28/2010
Solids, total suspended	Daily Maximum	37.5	75.8	lb/d	001G	02/28/2010
Solids, total suspended	Weekly Average	33.	36.9	lb/d	001G	02/28/2010
Total Ammonia Nitrogen	Daily Maximum	4.	4.98	mg/L	001G	02/28/2010

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Total Ammonia Nitrogen	Daily Maximum	3.3	3.85	lb/d	001G	02/28/2010
Solids, total suspended	Monthly Avg. Min.	30.	27.2	mg/L	001G	03/31/2010
Solids, total suspended	Daily Maximum	45.	73.	mg/L	001G	03/31/2010
Solids, total suspended	Daily Maximum	37.5	85.1	lb/d	001G	03/31/2010
Solids, settleable	Daily Maximum	1.	1.5	mL/L	001G	03/31/2010
Total Ammonia Nitrogen	Monthly Average	1.9	3.05	mg/L	001G	03/31/2010
Total Ammonia Nitrogen	Weekly Average	2.4	8.14	mg/L	001G	03/31/2010
Total Ammonia Nitrogen	Daily Maximum	4.	9.91	mg/L	001G	03/31/2010
Total Ammonia Nitrogen	Monthly Average	1.6	1.91	lb/d	001G	03/31/2010
Total Ammonia Nitrogen	Daily Maximum	3.3	6.33	lb/d	001G	03/31/2010
Solids, total suspended	Weekly Average	40.	44.2	mg/L	001G	04/30/2010
Solids, total suspended	Daily Maximum	45.	157.	mg/L	001G	04/30/2010
Solids, total suspended	Monthly Average	25.	35.7	lb/d	001G	04/30/2010
Solids, total suspended	Daily Maximum	37.5	140.5	lb/d	001G	04/30/2010
Solids, total suspended	Weekly Average	33.	108.3	lb/d	001G	04/30/2010
Solids, settleable	Daily Maximum	1.	4.5	mL/L	001G	04/30/2010
Total Ammonia Nitrogen	Weekly Average	2.4	2.63	mg/L	001G	04/30/2010
Total Ammonia Nitrogen	Daily Maximum	4.	5.66	mg/L	001G	04/30/2010
Total Ammonia Nitrogen	Daily Maximum	3.3	6.68	lb/d	001G	04/30/2010
E. coli, MTEC-MF	Monthly Geomean	126.	204.	#/100mL	001G	04/30/2010
E. coli, MTEC-MF	Daily Maximum	941.	2419.	#/100mL	001G	04/30/2010
BOD, carbonaceous, 05 day, 20 C	Monthly Average	10.	10.5	mg/L	001G	04/30/2010
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	20.	27.	mg/L	001G	04/30/2010
BOD, carbonaceous, 05 day, 20 C	Monthly Average	8.	8.6	lb/d	001G	04/30/2010
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	16.7	23.1	lb/d	001G	04/30/2010
BOD, carbonaceous, 05 day, 20 C	Weekly Average	13.	20.6	lb/d	001G	04/30/2010
Solids, total suspended	Monthly Avg. Min.	30.	7.3	mg/L	001G	05/31/2010
Total Ammonia Nitrogen	Monthly Average	1.1	5.67	mg/L	001G	05/31/2010
Total Ammonia Nitrogen	Weekly Average	2.	4.55	mg/L	001G	05/31/2010
Total Ammonia Nitrogen	Daily Maximum	2.5	28.	mg/L	001G	05/31/2010
Total Ammonia Nitrogen	Monthly Average	.9	3.71	lb/d	001G	05/31/2010
Total Ammonia Nitrogen	Daily Maximum	2.1	3.14	lb/d	001G	05/31/2010
Total Ammonia Nitrogen	Weekly Average	1.7	3.14	lb/d	001G	05/31/2010
E. coli, MTEC-MF	Monthly Geomean	126.	277.	#/100mL	001G	05/31/2010
E. coli, MTEC-MF	Daily Maximum	941.	2419.	#/100mL	001G	05/31/2010
Solids, total suspended	Monthly Avg. Min.	30.	13.9	mg/L	001G	06/30/2010
Solids, total suspended	Daily Maximum	45.	52.	mg/L	001G	06/30/2010
Total Ammonia Nitrogen	Monthly Average	1.1	14.73	mg/L	001G	06/30/2010
Total Ammonia Nitrogen	Weekly Average	2.	15.26	mg/L	001G	06/30/2010
Total Ammonia Nitrogen	Daily Maximum	2.5	43.01	mg/L	001G	06/30/2010
Total Ammonia Nitrogen	Monthly Average	.9	9.06	lb/d	001G	06/30/2010

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Total Ammonia Nitrogen	Daily Maximum	2.1	9.43	lb/d	001G	06/30/2010
Total Ammonia Nitrogen	Weekly Average	1.7	9.42	lb/d	001G	06/30/2010
E. coli, MTEC-MF	Daily Maximum	941.	2420.	#/100mL	001G	06/30/2010
Solids, total suspended	Monthly Avg. Min.	30.	9.5	mg/L	001G	07/31/2010
Solids, settleable	Daily Maximum	1.	4.5	mL/L	001G	07/31/2010
Total Ammonia Nitrogen	Monthly Average	1.1	6.35	mg/L	001G	07/31/2010
Total Ammonia Nitrogen	Weekly Average	2.	6.48	mg/L	001G	07/31/2010
Total Ammonia Nitrogen	Daily Maximum	2.5	18.82	mg/L	001G	07/31/2010
Total Ammonia Nitrogen	Monthly Average	.9	3.42	lb/d	001G	07/31/2010
Total Ammonia Nitrogen	Daily Maximum	2.1	3.5	lb/d	001G	07/31/2010
Total Ammonia Nitrogen	Weekly Average	1.7	17.3	lb/d	001G	07/31/2010
Solids, total suspended	Monthly Avg. Min.	30.	6.3	mg/L	001G	08/31/2010
Total Ammonia Nitrogen	Monthly Average	1.1	25.05	mg/L	001G	08/31/2010
Total Ammonia Nitrogen	Weekly Average	2.	25.05	mg/L	001G	08/31/2010
Total Ammonia Nitrogen	Daily Maximum	2.5	35.5	mġ/L	001G	08/31/2010
Total Ammonia Nitrogen	Monthly Average	.9	13.64	lb/d	001G	08/31/2010
Total Ammonia Nitrogen	Daily Maximum	2.1	13.64	lb/d	001G	08/31/2010
Total Ammonia Nitrogen	Weekly Average	1.7	13.64	lb/d	001G	08/31/2010
E. coli, MTEC-MF	Daily Maximum	941.	1733.	#/100mL	001G	08/31/2010
Solids, total suspended	Monthly Avg. Min.	30.	9.1	mg/L	001G	09/30/2010
Total Ammonia Nitrogen	Monthly Average	1.1	18.86	mg/L	001G	09/30/2010
Total Ammonia Nitrogen	Weekly Average	2.	21.45	mg/L	001G	09/30/2010
Total Ammonia Nitrogen	Daily Maximum	2.5	33.94	mg/L	001G	09/30/2010
Total Ammonia Nitrogen	Monthly Average	.9	8.22	lb/d	001G	09/30/2010
Total Ammonia Nitrogen	Daily Maximum	2.1	9.34	lb/d	001G	09/30/2010
Total Ammonia Nitrogen	Weekly Average	1.7	7.5	lb/d	001G	09/30/2010
E. coli, MTEC-MF	Monthly Geomean	126.	1448.	#/100mL	001G	09/30/2010
E. coli, MTEC-MF	Daily Maximum	941.	2420.	#/100mL	001G	09/30/2010
Solids, total suspended	Monthly Avg. Min.	30.	13.7	mg/L	001G	10/31/2010
Solids, settleable	Daily Maximum	1.	19.	mL/L	001G	10/31/2010
Total Ammonia Nitrogen	Monthly Average	1.1	4.59	mg/L	001G	10/31/2010
Total Ammonia Nitrogen	Weekly Average	2.	4.16	mg/L	001G	10/31/2010
Total Ammonia Nitrogen	Daily Maximum	2.5	10.19	mg/L	001G	10/31/2010
Total Ammonia Nitrogen	Monthly Average	.9	1.84	lb/d	001G	10/31/2010
E. coli, MTEC-MF	Monthly Geomean	126.	140.	#/100mL	001G	10/31/2010
Solids, total suspended	Monthly Avg. Min.	30.	17.7	mg/L	001G	11/30/2010
Solids, total suspended	Daily Maximum	45.	93.5	mg/L	001G	11/30/2010
Solids, total suspended	Daily Maximum	37.5	50.1	lb/d	001G	11/30/2010
Solids, settleable	Daily Maximum	1.	3.	mL/L	001G	11/30/2010
Total Ammonia Nitrogen	Monthly Average	1.9	14.81	mg/L	001G	11/30/2010
Total Ammonia Nitrogen	Weekly Average	2.4	23.22	mg/L	001G	11/30/2010
Total Ammonia Nitrogen	Daily Maximum	4.	23.91	mg/L	001G	11/30/2010
Total Ammonia Nitrogen	Monthly Average	1.6	9.91	lb/d	001G	11/30/2010
Total Ammonia Nitrogen	Daily Maximum	3.3	17.77	lb/d	001G	11/30/2010

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Total Ammonia Nitrogen	Weekly Average	2.	9.9	lb/d	001G	11/30/2010
E. coli, MTEC-MF	Monthly Geomean	126.	2056.	#/100mL	001G	11/30/2010
E. coli, MTEC-MF	Daily Maximum	941.	2420.	#/100mL	001G	11/30/2010
Solids, total suspended	Monthly Avg. Min.	30.	28.3	mg/L	001G	12/31/2010
Solids, total suspended	Daily Maximum	45.	133.	mg/L	001G	12/31/2010
Solids, total suspended	Daily Maximum	37.5	75.7	lb/d	001G	12/31/2010
Solids, settleable	Daily Maximum	1.	10.	mL/L	001G	12/31/2010
Total Ammonia Nitrogen	Monthly Average	1.9	4.3	mg/L	001G	12/31/2010
Total Ammonia Nitrogen	Weekly Average	2.4	6.31	mg/L	001G	12/31/2010
Total Ammonia Nitrogen	Daily Maximum	4.	12.1	mg/L	001G	12/31/2010
Total Ammonia Nitrogen	Monthly Average	1.6	2.31	lb/d	001G	12/31/2010
Total Ammonia Nitrogen	Daily Maximum	3.3	5.67	lb/d	001G	12/31/2010
E. coli, MTEC-MF	Daily Maximum	941.	2419.	#/100mL	001G	12/31/2010
Solids, total suspended	Monthly Avg. Min.	30.	5.9	mg/L	001G	01/31/2011
Solids, settleable	Daily Maximum	1.	1.5	mL/L	001G	01/31/2011
Total Ammonia Nitrogen	Monthly Average	1.9	2.9	mg/L	001G	01/31/2011
Total Ammonia Nitrogen	Weekly Average	2.4	11.93	mg/L	001G	01/31/2011
Total Ammonia Nitrogen	Daily Maximum	4.	6.22	mg/L	001G	01/31/2011
Total Ammonia Nitrogen	Monthly Average	1.6	1.86	lb/d	001G	01/31/2011
Total Ammonia Nitrogen	Daily Maximum	3.3	3.49	lb/d	001G	01/31/2011
Total Ammonia Nitrogen	Weekly Average	2.	2.6	lb/d	001G	01/31/2011
Solids, total suspended	Weekly Average	40.	72.5	mg/L	001G	02/28/2011
Solids, total suspended	Daily Maximum	45.	173.	mg/L	001G	02/28/2011
Solids, total suspended	Monthly Average	25.	55.1	lb/d	001G	02/28/2011
Solids, total suspended	Daily Maximum	37.5	129.9	lb/d	001G	02/28/2011
Solids, total suspended	Weekly Average	33.	102.4	lb/d	001G	02/28/2011
Solids, settleable	Daily Maximum	1.	5.	mL/L	001G	02/28/2011
Total Ammonia Nitrogen	Monthly Average	1.9	6.8	mg/L	001G	02/28/2011
Total Ammonia Nitrogen	Weekly Average	2.4	8.12	mg/L	001G	02/28/2011
Total Ammonia Nitrogen	Daily Maximum	4.	10.98	mg/L	001G	02/28/2011
Total Ammonia Nitrogen	Monthly Average	1.6	5.28	lb/d	001G	02/28/2011
Total Ammonia Nitrogen	Daily Maximum	3.3	8.24	lb/d	001G	02/28/2011
Total Ammonia Nitrogen	Weekly Average	2.	5.3	lb/d	001G	02/28/2011
E. coli, MTEC-MF	Monthly Geomean	126.	273.	#/100mL	001G	02/28/2011
E. coli, MTEC-MF	Daily Maximum	941.	2419.	#/100mL	001G	02/28/2011
Solids, total suspended	Daily Maximum	45.	162.	mg/L	001G	03/31/2011
Solids, total suspended	Monthly Average	25.	48.2	lb/d	001G	03/31/2011
Solids, total suspended	Daily Maximum	37.5	165.1	lb/d	001G	03/31/2011
Solids, total suspended	Weekly Average	33.	85.7	lb/d	001G	03/31/2011
Solids, settleable	Daily Maximum	1.	16.	mL/L	001G	03/31/2011
Total Ammonia Nitrogen	Daily Maximum	4.	4.14	mg/L	001G	03/31/2011
Total Ammonia Nitrogen	Daily Maximum	3.3	5.13	lb/d	001G	03/31/2011
E. coli, MTEC-MF	Daily Maximum	941.	961.	#/100mL	001G	03/31/2011
Solids, total suspended	Monthly Avg. Min.	30.	16.9	mg/L	001G	04/30/2011

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Solids, total suspended	Daily Maximum	45.	81.	mg/L	001G	04/30/2011
Solids, total suspended	Daily Maximum	37.5	65.6	lb/d	001G	04/30/2011
Solids, total suspended	Weekly Average	33.	82.5	lb/d	001G	04/30/2011
Solids, settleable	Daily Maximum	1.	1.5	mL/L	001G	04/30/2011
Total Ammonia Nitrogen	Monthly Average	1.9	2.39	mg/L	001G	04/30/2011
Total Ammonia Nitrogen	Weekly Average	2.4	6.42	mg/L	001G	04/30/2011
Total Ammonia Nitrogen	Daily Maximum	4.	8.18	mg/L	001G	04/30/2011
Total Ammonia Nitrogen	Monthly Average	1.6	2.62	lb/d	001G	04/30/2011
Total Ammonia Nitrogen	Daily Maximum	3.3	8.49	lb/d	001G	04/30/2011
Total Ammonia Nitrogen	Weekly Average	2.	2.3	lb/d	001G	04/30/2011
E. coli, MTEC-MF	Monthly Geomean	126.	134.	#/100mL	001G	04/30/2011
E. coli, MTEC-MF	Daily Maximum	941.	1413.6	#/100mL	001G	04/30/2011
Solids, total suspended	Monthly Avg. Min.	30.	20.4	mg/L	001G	05/31/2011
Solids, total suspended	Daily Maximum	45.	141.	mg/L	001G	05/31/2011
Solids, total suspended	Daily Maximum	37.5	90.7	lb/d	001G	05/31/2011
Solids, settleable	Daily Maximum	1.	4.	mL/L	001G	05/31/2011
Total Ammonia Nitrogen	Monthly Average	1.1	3.27	mg/L	001G	05/31/2011
Total Ammonia Nitrogen	Weekly Average	2.	3.27	mg/L	001G	05/31/2011
Total Ammonia Nitrogen	Daily Maximum	2.5	13.1	mg/L	001G	05/31/2011
Total Ammonia Nitrogen	Monthly Average	.9	3.41	lb/d	001G	05/31/2011
Total Ammonia Nitrogen	Daily Maximum	2.1	3.41	lb/d	001G	05/31/2011
Total Ammonia Nitrogen	Weekly Average	1.7	3.41	Ib/d	001G	05/31/2011
Solids, total suspended	Monthly Avg. Min.	30.	8.5	mg/L	001G	06/30/2011
Total Ammonia Nitrogen	Monthly Average	1.1	5.71	mg/L	001G	06/30/2011
Total Ammonia Nitrogen	Weekly Average	2.	5.24	mg/L	001G	06/30/2011
Total Ammonia Nitrogen	Daily Maximum	2.5	24.42	mg/L	001G	06/30/2011
Total Ammonia Nitrogen	Monthly Average	.9	3.67	lb/d	001G	06/30/2011
Total Ammonia Nitrogen	Daily Maximum	2.1	3.75	lb/d	001G	06/30/2011
Total Ammonia Nitrogen	Weekly Average	1.7	3.75	lb/d	001G	06/30/2011
E. coli, MTEC-MF	Monthly Geomean	126.	852.	#/100mL	001G	06/30/2011
E. coli, MTEC-MF	Daily Maximum	941.	2419.	#/100mL	001G	06/30/2011
Solids, total suspended	Monthly Avg. Min.	30.	11.	mg/L	001G	07/31/2011
Total Ammonia Nitrogen	Monthly Average	1.1	15.18	mg/L	001G	07/31/2011
Total Ammonia Nitrogen	Weekly Average	2.	14.3	mg/L	001G	07/31/2011
Total Ammonia Nitrogen	Daily Maximum	2.5	35.06	mg/L	001G	07/31/2011
Total Ammonia Nitrogen	Monthly Average	.9	8.1	lb/d	001G	07/31/2011
Total Ammonia Nitrogen	Daily Maximum	2.1	7.47	lb/d	001G	07/31/2011
Total Ammonia Nitrogen	Weekly Average	1.7	7.47	lb/d	001G	07/31/2011
E. coli, MTEC-MF	Monthly Geomean	126.	993.	#/100mL	001G	07/31/2011
E. coli, MTEC-MF	Daily Maximum	941.	72419.	#/100mL	001G	07/31/2011
Solids, total suspended	Monthly Avg. Min.	30.	9.	mg/L	001G	08/31/2011
Total Ammonia Nitrogen	Monthly Average	1.1	31.64	mg/L	001G	08/31/2011
Total Ammonia Nitrogen	Weekly Average	2.	32.97	mg/L	001G	08/31/2011
Total Ammonia Nitrogen	Daily Maximum	2.5	40.99	mg/L	001G	08/31/2011

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Total Ammonia Nitrogen	Monthly Average	.9	14.88	lb/d	001G	08/31/2011
Total Ammonia Nitrogen	Daily Maximum	2.1	15.6	lb/d	001G	08/31/2011
Total Ammonia Nitrogen	Weekly Average	1.7	15.6	lb/d	001G	08/31/2011
E. coli, MTEC-MF	Monthly Geomean	126.	1622.	#/100mL	001G	08/31/2011
E. coli, MTEC-MF	Daily Maximum	941.	2419.	#/100mL	001G	08/31/2011
Solids, total suspended	Monthly Avg. Min.	30.	5.6	mg/L	001G	09/30/2011
Total Ammonia Nitrogen	Monthly Average	1.1	6.52	mg/L	001G	09/30/2011
Total Ammonia Nitrogen	Weekly Average	2.	7.13	mg/L	001G	09/30/2011
Total Ammonia Nitrogen	Daily Maximum	2.5	20.5	mg/L	001G	09/30/2011
Total Ammonia Nitrogen	Monthly Average	.9	3.9	lb/d	001G	09/30/2011
Total Ammonia Nitrogen	Daily Maximum	2.1	4.06	lb/d	001G	09/30/2011
Total Ammonia Nitrogen	Weekly Average	1.7	4.06	lb/d	001G	09/30/2011
E. coli, MTEC-MF	Daily Maximum	941.	1011.	#/100mL	001G	09/30/2011
Solids, total suspended	Monthly Avg. Min.	30.	6.2	mg/L	001G	10/31/2011
Solids, settleable	Daily Maximum	1.	17.	mL/L	001G	10/31/2011
Total Ammonia Nitrogen	Monthly Average	1.1	6.89	mg/L	001G	10/31/2011
Total Ammonia Nitrogen	Weekly Average	2.	6.78	mg/L	001G	10/31/2011
Total Ammonia Nitrogen	Daily Maximum	2.5	16.35	mg/L	001G	10/31/2011
Total Ammonia Nitrogen	Monthly Average	.9	3.26	lb/d	001G	10/31/2011
Total Ammonia Nitrogen	Daily Maximum	2.1	3.23	lb/d	001G	10/31/2011
Total Ammonia Nitrogen	Weekly Average	1.7	3.23	lb/d	001G	10/31/2011
Solids, total suspended	Monthly Avg. Min.	30.	6.1	mg/L	001G	11/30/2011
Total Ammonia Nitrogen	Monthly Average	1.9	7.31	mg/L	001G	11/30/2011
Total Ammonia Nitrogen	Weekly Average	2.4	19.73	mg/L	001G	11/30/2011
Total Ammonia Nitrogen	Daily Maximum	4.	22.12	mg/L	001G	11/30/2011
Total Ammonia Nitrogen	Monthly Average	1.6	5.06	lb/d	001G	11/30/2011
Total Ammonia Nitrogen	Daily Maximum	3.3	15.55	lb/d	001G	11/30/2011
Total Ammonia Nitrogen	Weekly Average	2.	5.5	lb/d	001G	11/30/2011
E. coli, MTEC-MF	Monthly Geomean	126.	347.	#/100mL	001G	11/30/2011
E. coli, MTEC-MF	Daily Maximum	941.	2419.	#/100mL	001G	11/30/2011
Solids, total suspended	Monthly Avg. Min.	30.	22.3	mg/L	001G	12/31/2011
Solids, total suspended	Daily Maximum	45.	54.	mg/L	001G	12/31/2011
Solids, total suspended	Monthly Average	25.	31.2	lb/d	001G	12/31/2011
Solids, total suspended	Daily Maximum	37.5	76.2	lb/d	001G	12/31/2011
Solids, total suspended	Weekly Average	33.	71.6	lb/d	001G	12/31/2011
Solids, settleable	Daily Maximum	1.	9.	mL/L	001G	12/31/2011
BOD, carbonaceous, 05 day, 20 C	Monthly Average	8.	10.1	lb/d	001G	12/31/2011
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	16.7	21.8	lb/d	001G	12/31/2011
BOD, carbonaceous, 05 day, 20 C	Weekly Average	13.	18.2	lb/d	001G	12/31/2011
Solids, total suspended	Monthly Avg. Min.	30.	8.9	mg/L	001G	01/31/2012
Solids, total suspended	Daily Maximum	45.	55.	mg/L	001G	01/31/2012
Solids, total suspended	Daily Maximum	37.5	55.1	lb/d	001G	01/31/2012

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Solids, settleable	Daily Maximum	1.	16.	mL/L	001G	01/31/2012
Total Ammonia Nitrogen	Monthly Average	1.9	2.36	mg/L	001G	01/31/2012
Total Ammonia Nitrogen	Weekly Average	2.4	5.13	mg/L	001G	01/31/2012
Total Ammonia Nitrogen	Daily Maximum	4.	11.14	mg/L	001G	01/31/2012
Total Ammonia Nitrogen	Monthly Average	1.6	2.09	lb/d	001G	01/31/2012
Total Ammonia Nitrogen	Daily Maximum	3.3	9.6	lb/d	001G	01/31/2012
Total Ammonia Nitrogen	Weekly Average	2.	2.1	lb/d	001G	01/31/2012
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	16.7	17.6	lb/d	001G	01/31/2012
Solids, total suspended	Monthly Avg. Min.	30.	6.1	mg/L	001G	02/29/2012
Solids, settleable	Daily Maximum	1.	7.	mL/L	001G	02/29/2012
Solids, total suspended	Monthly Avg. Min.	30.	11.4	mg/L	001G	03/31/2012
Total Ammonia Nitrogen	Monthly Average	1.9	8.7	mg/L	001G	03/31/2012
Total Ammonia Nitrogen	Weekly Average	2.4	20.65	mg/L	001G	03/31/2012
Total Ammonia Nitrogen	Daily Maximum	4.	23.13	mg/L	001G	03/31/2012
Total Ammonia Nitrogen	Monthly Average	1.6	6.3	lb/d	001G	03/31/2012
Total Ammonia Nitrogen	Daily Maximum	3.3	21.07	lb/d	001G	03/31/2012
Total Ammonia Nitrogen	Weekly Average	2.	6.1	lb/d	001G	03/31/2012
Solids, total suspended	Monthly Avg. Min.	30.	7.4	mg/L	001G	04/30/2012
Total Ammonia Nitrogen	Monthly Average	1.9	14.6	mg/L	001G	04/30/2012
Total Ammonia Nitrogen	Weekly Average	2.4	21.91	mg/L	001G	04/30/2012
Total Ammonia Nitrogen	Daily Maximum	4.	25.42	mg/L	001G	04/30/2012
Total Ammonia Nitrogen	Monthly Average	1.6	7.48	lb/d	001G	04/30/2012
Total Ammonia Nitrogen	Daily Maximum	3.3	14.84	lb/d	001G	04/30/2012
Total Ammonia Nitrogen	Weekly Average	2.	7.5	lb/d	001G	04/30/2012
Solids, total suspended	Monthly Avg. Min.	30.	7.4	mg/L	001G	05/31/2012
Solids, settleable	Daily Maximum	1.	3.	mL/L	001G	05/31/2012
Total Ammonia Nitrogen	Monthly Average	1.1	16.39	mg/L	001G	05/31/2012
Total Ammonia Nitrogen	Weekly Average	2.	18.01	mg/L	001G	05/31/2012
Total Ammonia Nitrogen	Daily Maximum	2.5	25.87	mg/L	001G	05/31/2012
Total Ammonia Nitrogen	Monthly Average	.9	7.93	1b/d	001G	05/31/2012
Total Ammonia Nitrogen	Daily Maximum	2.1	8.77	lb/d	001G	05/31/2012
Total Ammonia Nitrogen	Weekly Average	1.7	8.77	lb/d	001G	05/31/2012
E. coli, MTEC-MF	Monthly Geomean	126.	363.	#/100mL	001G	05/31/2012
E. coli, MTEC-MF	Daily Maximum	941.	2119.6	#/100mL	001G	05/31/2012
Solids, total suspended	Monthly Avg. Min.	30.	6.	mg/L	001G	06/30/2012
Total Ammonia Nitrogen	Monthly Average	1.1	6.53	mg/L	001G	06/30/2012
Total Ammonia Nitrogen	Weekly Average	2.	6.55	mg/L	001G	06/30/2012
Total Ammonia Nitrogen	Daily Maximum	2.5	12.88	mg/L	001G	06/30/2012
Total Ammonia Nitrogen	Monthly Average	.9	2.91	lb/d	001G	06/30/2012
Total Ammonia Nitrogen	Daily Maximum	2.1	2.91	lb/d	001G	06/30/2012
Total Ammonia Nitrogen	Weekly Average	1.7	2.91	lb/d	001G	06/30/2012
E. coli, MTEC-MF	Monthly Geomean	126.	549.	#/100mL	001G	06/30/2012
E. coli, MTEC-MF	Daily Maximum	941.	2420.	#/100mL	001G	06/30/2012
Solids, total suspended	Monthly Avg. Min.	30.	3.9	mg/L	001G	07/31/2012

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Total Ammonia Nitrogen	Monthly Average	1.1	5.92	mg/L	001G	07/31/2012
Total Ammonia Nitrogen	Weekly Average	2.	5.92	mg/L	001G	07/31/2012
Total Ammonia Nitrogen	Daily Maximum	2.5	9.07	mg/L	001G	07/31/2012
Total Ammonia Nitrogen	Monthly Average	.9	3.24	lb/d	001G	07/31/2012
Total Ammonia Nitrogen	Daily Maximum	2.1	3.24	lb/d	001G	07/31/2012
Total Ammonia Nitrogen	Weekly Average	1.7	3.24	lb/d	001G	07/31/2012
E. coli, MTEC-MF	Monthly Geomean	126.	315.	#/100mL	001G	07/31/2012
E. coli, MTEC-MF	Daily Maximum	941.	1986.	#/100mL	001G	07/31/2012
Solids, total suspended	Monthly Avg. Min.	30.	9.1	mg/L	001G	08/31/2012
Solids, total suspended	Weekly Average	33.	86.	lb/d	001G	08/31/2012
Solids, settleable	Daily Maximum	1.	2.	mL/L	001G	08/31/2012
Total Ammonia Nitrogen	Monthly Average	1.1	3.47	mg/L	001G	08/31/2012
Total Ammonia Nitrogen	Weekly Average	2.	3.92	mg/L	001G	08/31/2012
Total Ammonia Nitrogen	Daily Maximum	2.5	6.83	mg/L	001G	08/31/2012
Total Ammonia Nitrogen	Monthly Average	.9	1.68	lb/d	001G	08/31/2012
E. coli, MTEC-MF	Monthly Geomean	126.	630.	#/100mL	001G	08/31/2012
E. coli, MTEC-MF	Daily Maximum	941.	2419.6	#/100mL	001G	08/31/2012
Solids, total suspended	Monthly Avg. Min.	30.	9.	mg/L	001G	09/30/2012
Total Ammonia Nitrogen	Monthly Average	1.1	1.82	mg/L	001G	09/30/2012
Total Ammonia Nitrogen	Weekly Average	2.	2.25	mg/L	001G	09/30/2012
Total Ammonia Nitrogen	Daily Maximum	2.5	5.26	mg/L	001G	09/30/2012
Total Ammonia Nitrogen	Monthly Average	.9	1.15	lb/d	001G	09/30/2012
E. coli, MTEC-MF	Monthly Geomean	126.	207.	#/100mL	001G	09/30/2012
E. coli, MTEC-MF	Daily Maximum	941.	980.	#/100mL	001G	09/30/2012
Solids, total suspended	Monthly Avg. Min.	30.	21.2	mg/L	001G	10/31/2012
Solids, total suspended	Daily Maximum	45.	73.	mg/L	001G	10/31/2012
Solids, total suspended	Daily Maximum	37.5	44.6	lb/d	001G	10/31/2012
Solids, settleable	Daily Maximum	1.	2.5	mL/L	001G	10/31/2012
Total Ammonia Nitrogen	Monthly Average	1.1	2.07	mg/L	001G	10/31/2012
Total Ammonia Nitrogen	Weekly Average	2.	2.15	mg/L	001G	10/31/2012
Total Ammonia Nitrogen	Daily Maximum	2.5	8.57	mg/L	001G	10/31/2012
Total Ammonia Nitrogen	Monthly Average	.9	1.31	lb/d	001G	10/31/2012
E. coli, MTEC-MF	Monthly Geomean	126.	1336.	#/100mL	001G	10/31/2012
E. coli, MTEC-MF	Daily Maximum	941.	2419.6	#/100mL	001G	10/31/2012
Solids, total suspended	Weekly Average	40.	40.7	mg/L	001G	11/30/2012
Solids, total suspended	Daily Maximum	45.	146.	mg/L	001G	11/30/2012
Solids, total suspended	Daily Maximum	37.5	66.5	lb/d	001G	11/30/2012
Solids, total suspended	Weekly Average	33.	54.9	lb/d	001G	11/30/2012
Solids, settleable	Daily Maximum	1.	19.5	mL/L	001G	11/30/2012
E. coli, MTEC-MF	Monthly Geomean	126.	1284.	#/100mL	001G	11/30/2012
E. coli, MTEC-MF	Daily Maximum	941.	2419.6	#/100mL	001G	11/30/2012
Solids, total suspended	Monthly Avg. Min.	30.	27.3	mg/L	001G	12/31/2012
Solids, total suspended	Weekly Average	40.	46.9	mg/L	001G	12/31/2012
Solids, total suspended	Daily Maximum	45.	123.	mg/L	001G	12/31/2012

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Solids, total suspended	Daily Maximum	37.5	146.4	lb/d	001G	12/31/2012
Solids, total suspended	Weekly Average	33.	55.1	lb/d⊸	001G	12/31/2012
Solids, settleable	Daily Maximum	1.	19.	mL/L	001G	12/31/2012
Total Ammonia Nitrogen	Weekly Average	2.4	3.25	mg/L	001G	12/31/2012
Total Ammonia Nitrogen	Daily Maximum	4.	6.5	mg/L	001G	12/31/2012
Total Ammonia Nitrogen	Daily Maximum	3.3	4.46	lb/d	001G	12/31/2012
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	16.7	21.2	lb/d	001G	12/31/2012
Solids, suspended percent removal	Monthly Avg. Min.	60.	54.4	%	001G	12/31/2012
Carbonaceous oxygen demand, % removal	Monthly Avg. Min.	75.	70.1	%	001G	12/31/2012
Solids, total suspended	Monthly Avg. Min.	30.	22.1	mg/L	001G	01/31/2013
Solids, total suspended	Daily Maximum	45.	106.	mg/L	001G	01/31/2013
Solids, total suspended	Monthly Average	25.	26.1	lb/d	001G	01/31/2013
Solids, total suspended	Daily Maximum	37.5	145.9	lb/d	001G	01/31/2013
Solids, total suspended	Weekly Average	33.	42.9	lb/d	001G	01/31/2013
Solids, settleable	Daily Maximum	1.	8.	mL/L	001G	01/31/2013
Total Ammonia Nitrogen	Weekly Average	2.4	2.57	mg/L	001G	01/31/2013
Total Ammonia Nitrogen	Daily Maximum	4.	6.94	mg/L	001G	01/31/2013
Total Ammonia Nitrogen	Monthly Average	1.6	2.2	lb/d	001G	01/31/2013
Total Ammonia Nitrogen	Daily Maximum	3.3	9.55	lb/d	001G	01/31/2013
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	16.7	27.5	lb/d	001G	01/31/2013
BOD, carbonaceous, 05 day, 20 C	Weekly Average	13.	13.8	lb/d	001G	01/31/2013
Solids, suspended percent removal	Monthly Avg. Min.	60.	49.5	%	001G	01/31/2013
Solids, total suspended	Monthly Avg. Min.	30.	4.7	mg/L	001G	02/28/2013
Total Ammonia Nitrogen	Monthly Average	1.9	6.19	mg/L	001G	02/28/2013
Total Ammonia Nitrogen	Weekly Average	2.4	8.85	mg/L	001G	02/28/2013
Total Ammonia Nitrogen	Daily Maximum	4.	9.86	mg/L	001G	02/28/2013
Total Ammonia Nitrogen	Monthly Average	1.6	4.37	lb/d	001G	02/28/2013
Total Ammonia Nitrogen	Daily Maximum	3.3	7.15	lb/d	001G	02/28/2013
Total Ammonia Nitrogen	Weekly Average	2.	4.9	lb/d	001G	02/28/2013
E. coli, MTEC-MF	Daily Maximum	941.	1046.2	#/100mL	001G	02/28/2013
BOD, carbonaceous, 05 day, 20 C	Weekly Average	13.	13.1	lb/d	001G	02/28/2013
Solids, total suspended	Monthly Avg. Min.	30.	4.5	mg/L	001G	03/31/2013
Total Ammonia Nitrogen	Monthly Average	1.9	3.3	mg/L	001G	03/31/2013
Total Ammonia Nitrogen	Weekly Average	2.4	10.3	mg/L	001G	03/31/2013
Total Ammonia Nitrogen	Daily Maximum	4.	12.3	mg/L	001G	03/31/2013
Total Ammonia Nitrogen	Monthly Average	1.6	2.44	lb/d .	001G	03/31/2013
Total Ammonia Nitrogen	Daily Maximum	3.3	9.97	lb/d	001G	03/31/2013
Solids, total suspended	Monthly Avg. Min.	30.	5.2	mg/L	001G	04/30/2013
Total Ammonia Nitrogen	Monthly Average	1.9	2.72	mg/L	001G	04/30/2013
Total Ammonia Nitrogen	Weekly Average	2.4	2.72	mg/L	001G	04/30/2013

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Total Ammonia Nitrogen	Daily Maximum	4.	8.06	mg/L	001G	04/30/2013
Total Ammonia Nitrogen	Monthly Average	1.6	2.4	lb/d	001G	04/30/2013
Total Ammonia Nitrogen	Daily Maximum	3.3	7.27	lb/d	001G	04/30/2013
Total Ammonia Nitrogen	Weekly Average	2.	5.4	lb/d	001G	04/30/2013
Solids, total suspended	Monthly Avg. Min.	30.	14.3	mg/L	001G	05/31/2013
Solids, total suspended	Daily Maximum	37.5	46.5	lb/d	001G	05/31/2013
Solids, settleable	Daily Maximum	1.	15.	mL/L	001G	05/31/2013
Total Ammonia Nitrogen	Monthly Average	1.1	3.93	mg/L	001G	05/31/2013
Total Ammonia Nitrogen	Weekly Average	2.	3.22	mg/L	001G	05/31/2013
Total Ammonia Nitrogen	Daily Maximum	2.5	3.93	mg/L	001G	05/31/2013
Total Ammonia Nitrogen	Monthly Average	.9	2.82	lb/d	001G	05/31/2013
Total Ammonia Nitrogen	Daily Maximum	2.1	2.24	lb/d	001G	05/31/2013
Solids, suspended percent removal	Monthly Avg. Min.	60.	58.9	%	001G	05/31/2013
Solids, total suspended	Monthly Avg. Min.	30.	5.1	mg/L	001G	06/30/2013
Total Ammonia Nitrogen	Monthly Average	1.1	3.84	mg/L	001G	06/30/2013
Total Ammonia Nitrogen	Weekly Average	2.	5.4	mg/L	001G	06/30/2013
Total Ammonia Nitrogen	Daily Maximum	2.5	9.69	mg/L	001G	06/30/2013
Total Ammonia Nitrogen	Monthly Average	.9	2.27	lb/d	001G	06/30/2013
Total Ammonia Nitrogen	Daily Maximum	2.1	2.84	lb/d	001G	06/30/2013
Total Ammonia Nitrogen	Weekly Average	1.7	2.84	lb/d	001G	06/30/2013
Solids, total suspended	Monthly Avg. Min.	30.	9.5	mg/L	001G	07/31/2013
Solids, total suspended	Daily Maximum	37.5	56.4	lb/d	001G	07/31/2013
Solids, settleable	Daily Maximum	1.	2.	mL/L	001G	07/31/2013
Total Ammonia Nitrogen	Monthly Average	1.1	4.32	mg/L	001G	07/31/2013
Total Ammonia Nitrogen	Weekly Average	2.	3.88	mg/L	001G	07/31/2013
Total Ammonia Nitrogen	Daily Maximum	2.5	12.38	mg/L	001G	07/31/2013
Total Ammonia Nitrogen	Monthly Average	.9	3.16	lb/d	001G	07/31/2013
Total Ammonia Nitrogen	Daily Maximum	2.1	2.8	lb/d	001G	07/31/2013
Total Ammonia Nitrogen	Weekly Average	1.7	2.8	lb/d	001G	07/31/2013
E. coli, MTEC-MF	Daily Maximum	941.	2419.6	#/100mL	001G	07/31/2013
Solids, total suspended	Monthly Avg. Min.	30.	3.8	mg/L	001G	08/31/2013
Total Ammonia Nitrogen	Monthly Average	1.1	8.56	mg/L	001G	08/31/2013
Total Ammonia Nitrogen	Weekly Average	2.	8.98	mg/L	001G	08/31/2013
Total Ammonia Nitrogen	Daily Maximum	2.5	19.5	mg/L	001G	08/31/2013
Total Ammonia Nitrogen	Monthly Average	.9	6.92	lb/d	001G	08/31/2013
Total Ammonia Nitrogen	Daily Maximum	2.1	16.4	lb/d	001G	08/31/2013
Total Ammonia Nitrogen	Weekly Average	1.7	7.01	lb/d	001G	08/31/2013
E. coli, MTEC-MF	Daily Maximum	941.	1986.3	#/100mL	001G	08/31/2013
Solids, total suspended	Monthly Avg. Min.	30.	4.	mg/L	001G	09/30/2013
Total Ammonia Nitrogen	Monthly Average	1.1	14.42	mg/L	001G	09/30/2013
Total Ammonia Nitrogen	Weekly Average	2.	14.42	mg/L	001G	09/30/2013
Total Ammonia Nitrogen	Daily Maximum	2.5	22.1	mg/L	001G	09/30/2013
Total Ammonia Nitrogen	Monthly Average	.9	9.65	lb/d	001G	09/30/2013
Total Ammonia Nitrogen	Daily Maximum	2.1	14.7	lb/d	001G	09/30/2013

Parameter Description	Violation	Limit Value	DMR Value	Units	Outfall	Reporting Period
Total Ammonia Nitrogen	Weekly Average	1.7	9.65	lb/d	001G	09/30/2013
Solids, total suspended	Monthly Avg. Min.	30.	3.	mg/L	001G	10/31/2013
Total Ammonia Nitrogen	Monthly Average	1.1	14.2	mg/L	001G	10/31/2013
Total Ammonia Nitrogen	Weekly Average	2.	14.2	mg/L	001G	10/31/2013
Total Ammonia Nitrogen	Daily Maximum	2.5	29.2	mg/L	001G	10/31/2013
Total Ammonia Nitrogen	Monthly Average	.9	9.7	lb/d	001G	10/31/2013
Total Ammonia Nitrogen	Daily Maximum	2.1	20.6	lb/d	001G	10/31/2013
Total Ammonia Nitrogen	Weekly Average	1.7	8.2	lb/d	001G	10/31/2013
Solids, total suspended	Weekly Average	40.	105.3	mg/L	001G	11/30/2013
Solids, total suspended	Daily Maximum	45.	378.5	mg/L	001G	11/30/2013
Solids, total suspended	Monthly Average	25.	100.4	lb/d	001G	11/30/2013
Solids, total suspended	Daily Maximum	37.5	419.2	lb/d	001G	11/30/2013
Solids, total suspended	Weekly Average	33.	294.8	lb/d	001G	11/30/2013
Solids, settleable	Daily Maximum	1.	70.	mL/L	001G	11/30/2013
Total Ammonia Nitrogen	Monthly Average	1.9	10.5	mg/L	001G	11/30/2013
Total Ammonia Nitrogen	Weekly Average	2.4	12.8	mg/L	001G	11/30/2013
Total Ammonia Nitrogen	Daily Maximum	4.	24.5	mg/L	001G	11/30/2013
Total Ammonia Nitrogen	Monthly Average	1.6	6.09	lb/d	001G	11/30/2013
Total Ammonia Nitrogen	Daily Maximum	3.3	20.73	lb/d	001G	11/30/2013
Total Ammonia Nitrogen	Weekly Average	2.	7.7	lb/d	001G	11/30/2013
E. coli, MTEC-MF	Daily Maximum	941.	1732.9	#/100mL	001G	11/30/2013
BOD, carbonaceous, 05 day, 20 C	Monthly Average	10.	11.2	mg/L	001G	11/30/2013
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	20.	21.1	mg/L	001G	11/30/2013
BOD, carbonaceous, 05 day, 20 C	Daily Maximum	16.7	24.2	lb/d	001G	11/30/2013
BOD, carbonaceous, 05 day, 20 C	Weekly Average	13.	19.2	lb/d	001G	11/30/2013
Solids, suspended percent removal	Monthly Avg. Min.	60.	41.6	%	001G	11/30/2013

ENCLOSURE C

RIGHT TO ASSERT BUSINESS CONFIDENTIALITY CLAIMS (40 C.F.R. Part 2)

Except for effluent data, you may, if you desire, assert a business confidentiality claim as to any or all of the information that the EPA is requesting from you. The EPA regulation relating to business confidentiality claims is found at 40 C.F.R. Part 2.

If you assert such a claim for the requested information, the EPA will only disclose the information to the extent and under the procedures set out in the cited regulations. If no business confidentiality claim accompanies the information, the EPA may make the information available to the public without any further notice to you.

40 C.F.R. § 2.203(b). Method and time of asserting business confidentiality claim. A business which is submitting information to the EPA may assert a business confidentiality claim covering the information by placing on (or attaching to) the information, at the time it is submitted to the EPA, a cover sheet, stamped or typed legend, or other suitable form of notice employing language such as trade secret, proprietary, or company confidential. Allegedly confidential portions of otherwise non-confidential documents should be clearly identified by the business, and may be submitted separately to facilitate identification and handling by the EPA. If the business desires confidential treatment only until a certain date or until the occurrence of a certain event, the notice should so state.

United States Environmental Protection Agency Region 4

Science and Ecosystem Support Division 980 College Station Road Athens, Georgia 30605-2720



Diagnostic Evaluation Report

City of Cornersville Sewage Treatment Plant Cornersville, Tennessee Inspection Date: May 6-7, 2013 NPDES Permit No. TN0061841 SESD Project ID No. 13-0346

Requestor: Maurice Horsey, Chief
Municipal and Industrial Enforcement Section
Water Protection Division
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SESD Project Leader: Jairo Castillo, P.E. Environmental Engineer Enforcement Section U.S. EPA Region 4 980 College Station Road Athens, Georgia 30605-2720

Title and Approval Sheet

Title: Diagnostic Evaluation Inspection Report
City of Cornersville Sewage Treatment Plant (STP)

Approving Official:

Mike Bowden, Chief Enforcement Section

Enforcement and Investigations Branch

Date

SESD Project Leader:

Jairo Castillo, PE, Environmental Engineer

Enforcement Section

Enforcement and Investigations Branch

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Diagnostic Evaluation Report City of Cornersville Sewage Treatment Plant Cornersville, Tennessee- No. TN0061841

EXECUTIVE SUMMARY

During the week of May 6, 2013, representatives of the U.S. Environmental Protection Agency, Science and Ecosystem Support Division (USEPA-SESD), conducted a Diagnostic Evaluation at the Cornersville Sewage Treatment Plant (STP) in Cornersville, Tennessee. The purpose of the study was to assess the overall operation of the Cornersville STP. The facility is a 0.1 MGD that serves residential communities and commercial properties. The plant was built in 1990 and consists of one 1.2 million gallon equalization tank with three mechanical aerators, followed by two sequencing batch reactors (SBRs) tanks, an ultraviolet (UV) disinfection system, and an effluent cascade aerator channel that discharges into Town Creek (See Figure 1). Plant Discharge Monitoring Reports (DMRs) indicated numerous violations for ammonia -nitrogen, carbonaceous biochemical oxygen demand (CBOD₅), total suspended solids (TSS), *E. coli*, and settleable solids during 2012 and 2013. At the time of inspection, the facility was operating in storm mode due to a severe rain event that occurred within 48 hrs before the inspection. This report provides a description of the evaluation, including findings and recommendations of the inspection.

The diagnostic evaluation included the following tasks:

- Assessing the design, operations and management factors limiting treatment performance.
- Review of operations and management of the wastewater treatment plant.
- Characterization of the influent raw wastewater and final effluent.
- Evaluation of the unit processes performance via visual observations, sampling, data review, and process control testing.
- Determination of unit process operating parameters (e.g. Mean cell residence time, F/M, etc.).
- Evaluation of the NPDES self-monitoring program including sampling, flow measurement, records and reports, and laboratory procedures.

The major findings were as follows:

- The Cornersville STP aeration basin was operating with a mixed liquor suspended solids (MLSS) of 4,300 mg/l. This value was at the higher end of the recommended range (2000 to 5,000 mg/l) of MLSS in an SBR. The MCRT was 43 days. The MCRT exceeded the recommended value for optimal biological activity (up to 30 days) because of the high solids content in the SBR in relation to the amount of sludge wasted. The F/M value was 0.02 lbs BOD/day/lbs MLVSS, which was below the recommended range for proper CBOD₅ and ammonia as nitrogen removal (0.04 to 0.10 lbs BOD/day/lbs MLVSS).
- Low DO concentrations were found in the SBR react cycle. DO concentrations ranged from 0.28 to 1.15 mg/l.

- DMRs from 2012 and 2013 revealed numerous permit violations for ammonia-nitrogen, CBOD₅,
 TSS, E. coli, and Settleable Solids.
- Rising sludge was observed in the settlometer test due to denitrification. Low DO concentrations in the SBRs react cycle may result in dentrification during the decant cycle.
- The facility manually applied sodium bicarbonate to add alkalinity to the SBR biomass to control the pH. The average pH of the activated sludge from several instantaneous readings was 7. Slime and solids accumulation were observed on the UV lamp's surface. Slime buildup in the UV lamps and high suspended solids concentrations in wastewater adversely affect the UV system disinfection efficiency.
- The flow data observed on the DMRs shows that of the last fifteen months (Jan-2012 through Mar-2013), the effluent flow equaled or exceeded the design capacity of the plant (0.100 MGD) for five months. For the months of January and February, 2012, the average effluent flows were 0.108 mgd and 0.105 mgd, respectively. For the months of January and March, 2013, the average flows were 0.131 mgd and 0.098 mgd, respectively.
- An EPA instantaneous flow meter check revealed that the secondary flow meter measured 25
 percent below the instantaneous flow measured at the primary device (Parshall flume), which
 exceeded the EPA accepted accuracy range of ±10 percent.
- Solids buildup was observed on the discharge channel and the Parshall flume crest.
- No preliminary treatment was observed at the plant.
- The equalization tank was observed near to exceeding capacity. The tank overflows during severe rain events.
- The influent composite sampler tubing was clogged
- The influent automatic sampler was located after the equalization tank.
- The pH 7 buffer solution used for instrument calibration exceeded the expiration date (January, 2013).
- Transcription errors were observed in the 2012 and 2013 DMRs.

Recommendations:

- The Cornersville STP staff should focus on monitoring key operating parameters (MCRT, F/M, MLSS, sludge age) on a daily basis, interpret the data in relation to the effluent quality and sludge settleability, and conduct the necessary adjustments to meet the limits of the NPDES permit, specifically for CBOD₅, TSS, settleable solids, ammonia-nitrogen, and *E. coli* Bacteria. The solids inventory should be gradually reduced to obtain an MCRT that provides for adequate CBOD₅ removal and nitrification, as well as good settling characteristics. This will also bring the F/M ratio up, keeping the biological process more balanced and in an acceptable range of 0.4 to 0.10 lbs BOD/day-lbs MLVSS.
- Minimum concentrations of 1 to 3 mg/l are required in the SBR fill/react cycle to provide an optimal biological activity for CBOD₅ and ammonia-nitrogen biological removal. In addition, the F/M ratio of 0.02 should be increased to the acceptable range by carefully increasing sludge wasting until the treatment process improves.

- The permittee should consider installing an automatic lime or sodium bicarbonate feeders and pH probes in the SBRs units in order to maintain the activated sludge in the target range of 7.5 to 9.0 standard units.
- The solids inventory should be gradually reduced to obtain an MCRT that provides for adequate CBOD₅ removal and proper nitrification, as well as good settling characteristics.
- The permittee needs to provide proper maintenance (cleaning) to the UV system to improve the system operation and comply with the limits of the NPDES permit for E.Coli.
- The SBR process control testing should be increased to provide for optimal treatment. The following process control tests should be conducted regularly: aeration basin DO profile, Settlometer test, aeration basin pH, microscopic examination, and F/M calculations.
- The effluent secondary flow meter should be recalibrated to achieve accurate readings to within the EPA accepted accuracy range of +/- 10 percent. The upstream channel in the flume needs to be cleaned and properly maintained.
- The permittee should monitor the average influent flow of the plant to identify if the plant exceeds the 85 percent of the plant's design criteria on an average annual basis based on the previous twelve months of data. If the 85 percent of design criteria is exceeded, the permittee should provide a schedule to expand the plant's capacity to begin within one year of the exceedance.
- The permittee should consider the installation of a preliminary treatment process, such as screening and grit removal to reduce the inorganic solids in the equalization tank and subsequent units.
- The equalization tank should be evaluated for a potential accumulation of solids that could reduce its capacity. Additionally, the permittee should also assess the city's sewer system to detect potential infiltration problems that appear to increase the influent flow of the plant during rain events.
- The influent automatic sampler should be moved upstream of the equalization basin and should be properly maintained. The intake line should be checked regularly for rags and other debris that can cause clogging problems. Or, the line should be moved or repositioned in a location where rags and debris are not a problem.
- The permittee should assess the SBR diffuser system performance and operation including the blower capacity to determine if the system has the capability of increasing the air supply needed for the SBRs treatment process. A gradual reduction in the MLSS should result in lower oxygen requirements in the reactor.
- The SBR total cycle time should be increased to approximately five hours (from 4.35 hours) to provide the required time for proper nitrification (CSU, 2008).
- Transcription errors on the DMRs should be corrected and can be prevented with an in-house quality assurance/quality control process. DMRs should be routinely cross-checked and reviewed to avoid future errors.
- Additional O&M training for the staff would improve plant operations.

1. INTRODUCTION

During the week of May 6, 2013, representatives of the U.S. Environmental Protection Agency, Science and Ecosystem Support Division (USEPA – SESD), conducted a Diagnostic Evaluation (DE) at the City of Cornersville Sewage Treatment Plant (STP) at 1880 Ostella Rd Cornersville, Tennessee. The DE was performed at the request of the EPA Region 4 Water Protection Division, as the permittee has had difficulty meeting their NPDES permit limits.

The following personnel participated in the Diagnostic Evaluation:

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Dewitt Logsdon	TDEC, Inspector	(931) 490-3940
Kent Sweeton	Water and Wastewater Supervisor	(931) 359-6831
Denise Massey	Operator	(931) 359-2363

2. BACKGROUND

The City of Cornersville is in Marshall County Tennessee, approximately sixty miles south of Nashville. Since 2007, the City of Lewisburg, located approximately 6 miles north of Cornersville, owns and operates the plant through its Water and Wastewater Treatment Program. The community has a population of approximately 1,194 people. The Cornersville STP is a 0.1 MGD that serves residential communities and commercial properties. Previous Discharge Monitoring Reports (DMRs) indicated numerous violations for ammonia- nitrogen, CBOD₅, total suspended solids (TSS), *E. coli*, and settleable solids during 2012 and 2013. The recommendations in this report address the current plant as configured.

3. FACILITY SITE REVIEW

The Cornersville STP is a 0.1 MGD municipal wastewater treatment plant in Cornersville, TN. The plant was built in 1990. The plant operates 24 hours per day, 365 days per year and consists of a one 1.2 million gallon equalization tank with three mechanical aerators, followed by two sequencing batch reactors (SBRs) tanks in, and UV disinfection system. The sludge generated from the plant was treated in the Lewisburg Wastewater Treatment Plant. Figure 1 shows the facility's treatment processes and direction of the flow.



3.1. Permit

The Cornersville STP is authorized to discharge under the permit No. TN0061841. The NPDES permit issued by the Tennessee DEC, became effective on January 1, 2013 and expires on November 30, 2017. The name and description of the facility, the location of the outfall and the name of the receiving waters were as described in the permit.

3.2. Records and Reports

Self monitoring records consisted of the following:

- Discharge monitoring reports (DMRs)
- Monthly operating reports (MORs)
- *Laboratory bench sheets
- **■**Daily operational sheets
- Calibration records

The self monitoring records were kept for a minimum of three years. The permittee's self monitoring data for 2012 and 2013 are shown in Tables 1, 2, and 3. Numbers highlighted in "Red" and shaded indicate effluent violation of a listed permit parameter.

Deficiencies:

- Discharge Monitoring Reports (DMRs) indicated numerous violations for ammonia as nitrogen, total suspended solids (TSS), E. coli, and settleable solids on 2012 and 2013.
- Transcription errors were found in the spreadsheets and discharge monitoring reports were not

consistent with NPDES self-monitoring requirements regarding sampling documentation and reporting of parameter concentrations and loadings. The correct spreadsheet cell was not used for calculating the weekly average value. The spreadsheet cell was referenced to the ammonia daily maximum value (lbs/d) instead of the weekly average value.

Regulatory Requirement: 40CFR part 122.41 (a) (1) Duty to comply. The permittee shall comply with effluent standards or prohibitions established under section 307 (a) of the Clean Water Act for toxic pollutants and with standards for sewage sludge use or disposal established under section 405 (d) of the CWA within the time provided in the regulations that establish these standards or prohibitions or standards for sewage sludge use or disposal, even if the permit has not yet been modified to incorporate the requirement.

<u>Regulatory Requirement:</u> 40CFR part 122.41 (I) (4) Monitoring reports. Monitoring results shall be reported at the intervals specified elsewhere in this permit.

<u>Suggestion:</u> Peer review should be conducted before submitting the DMRs to ensure accuracy in reporting.

Table 1: Cornersville STP Self Monitoring Data (DMRs 2012-2013)-Solids

	Flow (mgd)	Inf. CBOD ₃ · (mg/L)		Eff. GBOD ₅ (mg/l)		CBOD ₅ Percent Removal		Eff. CBOD (lb/d)	
		MA	MA	WA	DM	MA	MA	WA	DM
Permit Limit	Hor	Report	10	15	20	75	8	13	16.7
Jan-2012	0.108	49.4	. 4.63	4.6	17.6	89	4.23	4.2	17.60
Feb-2012	0.105	37.7	3,23	3.2	4,4	92	2.82	2.8	3.9
Mar-2012	0.099	67.3	5.2	5.L	9.9	92	.4.0	4.1	7.0
Apr-2012	0.062	127.0	6.7	6.7	14.8	94.7	3.4	5.8	8,7
May-2012	0.062	L16.2	5.78	- 5.9	18.8	94.9	2.75	2.8	8
Jun-2012	0.057	112.9	3.89	3.41	6.9	97.1	1.71	1.8	3
Jul-2012	0.066	123.0	3.5	3.5	4.5	97.1	1.86	1.9	4
Aug-2012	0.058	105,7	5,5	8.4	8.4	94.7	2.39	2.5	4
Oct-2012	0.073	70.2	5.04	5.2	14.1	92.2	3.0	3.L	10
Nov-2012	0.052	114.9	8.4	8.3	19.5	92.3	3.6	8.0	8.5
Dec-2012	0.086	40.6	6.7	9.1	17.8	83	5.3	8.9	21.2
Jan-2013	0.131	32.7	6.6	5.7	20	81.3	7.7	13.8	27.5
Feb-2013	0.086	91.5	4.1	6.5	6.5	96,5	2.1	13.1	4.6
Mar-2013	0.098	61.7	2.8	3.0	5.3	94.2	2.3	3.2	4.7

Note: DM= Daily Maximum; MA= Monthly Average; WA= Weekly Average, No data provided for the month of Sept, 2012.

Table 2: Cornersville STP Self Monitoring Data (DMRs 2012-2013)-Bacteria and Solids

	E. Coli (W/100mL)		Inf Effluent TSS (mg/l) (mg/l)		TSS *Percent Removal		Effluent TSS (lb/d)			
	DM	Mean GM	MA	MA	WA	DM	MA	MA	WA	DM
Permit Limit	941	126		30	40	45	60	25	33	37.6
Jan-2012	866.4	7	49	8,9	8,9	55.0	89,6	8.3	24.5	55.1
Feb-2012	34	4	62	· 6.1	5.7	10.5	90.5	5.4	7.3	9.6
Mar-2012	78	7	82	11.4	11.4	19.0	86.2	8.9	11.0	12.4
Apr-2012	14	193.50	86	7.4	7,4	12.0	90.8	3.7	4.8	7.0
May-2012	363	2,419	178	7,4	5.4	33.0	91.2	. 7.93	3.4	14.6
Jun-2012	549	2420	87	6.0	7.8	12.88	96.0	6.0	7.8	9.0
Jul-2012	315	1986	105	3.9	3.9	6.0	96.0	2.0	2.3	4.0
Aug-2012	630	2419.6	77	9.1	9.9	33.0	87.2	4.3	8.6	16.8
Oct-2012	1336	2420	50	21.2	22.2	73.0	71.0	12.8	32.7	44.6
Nov-2012	1284	2419.6	74	41.8	40.7	146.0	68.9	8.1	54.9	66.5
Dec-2012	8	65.30	70	27.3	46.9	123.0	54.1	23.7	55.1	146.4
Jan-2013	33	139.60	39	22.1	16.1	106.0	49,5	26.1	42.9	145.9
Feb-2013	29	1046.20	86	4.7	5.L	12.5	94.0	3,4	11.5	8.8
Mar-2013	7	25.90	69 '	4.5	5.0	8.0	92.4	3.8	6,4	6,9

Note: DM= Daily Maximum, MA= Monthly Average, WA= Weekly Average, No data provided for the month of Sept, 2012. Influent Samples were collected after the equalization basin and do not reflect raw influent versus final effluent percent removals.

Table 3: Cornersville STP Self Monitoring Data (DMRs 2012-2013)-DO, pH, Solids, Nutrients

	Eff. DO	pH (S.U)	Settienble solids	Am	monia (NH ₃) (mg/L)	ns N	Ап	monia (NH ₃) as (lb/d))*	N	P (mg/L)
	(mg/l)		(ml/L)	MA	WA	DM	MA	WA	DM	OF THE REAL PROPERTY.
Permit Limit	Min 6	6-9	1	1.1/1.9	2/2.4	2.7/4.0	0.9/1.6	1.7/2.0	2.1/3.3	Report
Jan-2012	9.3	6,9/7,6	16.00	2.6	5.13	11.14	2.09	2,1	9.68	0.00
Feb-2012	9.6	7.2/7.8	7	0.22	0.5	1.29	0.2	0.5	1,5	0.00
Mar-2012	9.2	7.5/7.9	BDL	8,70	20.65	23,13	6.30	6.1	21.07	0,00
Apr-2012	7.7	7.0/7.9	BDL	14.60	21.91	25.42	7.48	7.5	14.3	0.00
May-2012	8,9	7.3/7.9	3	7,93	8.77	25.87	16.39	8.77	12.77	3.65
Jun-2012	8	6.8/7.7	0.40	2.91	2.91	2.91	6.53	6.55	12.8	6.25
Jul-2012	9.0	7.3/7.8	0.30	5.92	5.92	9.07	3.24	3.24	3.24	3,35
Aug-2012	9.2	7.5/7.9	2.0	3.47	3.92	6,83	1.68	1,44	1.44	6.90
Oct-2012	7.4	7.4/7.9	2.50	2.07	2.15	8.57	i 1.31	1.38	1,38	3.35
Nov-2012	9.0	6.9/7.8	19.50	0.98	2.10	3.19	0.42	0.4	1.35	0.00
Dec-2012	8.1	7.4/8.0	19.0	0.93	3.25	6.50	0.61	0.7	4.46	0.00
Jan-2013	9,9	7.4/7.7	8.0	1.81	2.57	6.94	2.20	1.3	9.55	0.00
Feb-2013	10.2	7.2/7.8	1.0	6.19	8.85	9.86	4.37	4.9	7.15	1.70
Mar-2013	10.3	7.4/7.7	BDL	3.30	10.30	12.30	2.44	2.0	9.97	0,00

**Error transcription in spreadsheet

Note: DM= Daily Maximum; MA= Monthly Average; WA= Weekly Average; No data provided for the month of Sept, 2012. Ammonia limits for Summer: OctApril/Winter: Nov-May)

3.3. Flow Measurement

The facility's influent flow was measured by an electromagnetic inline flow meter (MCCRO). The effluent flow was measured with a 6-inch Parshall flume and a secondary flow meter (STI 345 Magnetrol), which included a chart recorder and totalizer. The secondary flow meter was used for permit reporting. The effluent meter was last calibrated in February, 2013. Table 4 describes the results of the EPA check of the secondary flow measurement device for accuracy.

Table 4: Flow Measurement

Flow Device	Size	Instantaneous Head	Flow (gpm)	Error
Effluent Parshall Flume	6-in	0.6	412.5	25%
Plant Secondary Flow Meter	-	0.5	309.2	

Note: Eleven (11) sequencing discharges per day, 30 min duration

The EPA used a Teledyne ISCO Ultrasonic Flowmeter to measure the effluent flow to collect a flow proportional 24-hour composite sample. The EPA effluent flow was 0.185 mgd. During the same period, the effluent flow of the plant was 0.170 mgd; an 8.1 percent difference compared to the EPA flow value. In addition, the EPA effluent flow was 85 percent higher than the design flow of the treatment plant.

Solids buildup was observed in the effluent trough and in the Parshall flume crest. The solids accumulation in the Parshall flume crest appears to affect the accuracy of flow measurement, thus generating a reading difference of 25 percent between the primary and secondary flow measuring devices.

The flow data observed on the DMRs shows that for the last fifteen months (Jan-2012 through Mar-2013), the average monthly effluent flow equals or exceeds the design capacity of the plant (0.100 MGD) in five months. For the months of January and February, 2012, the monthly average effluent flows were 0.108 mgd and 0.105 mgd, respectively. For the months of January and March, 2013, the average flows were 0.131 mgd and 0.098 mgd, respectively. These values exceeded the 85 percent of the plant design criteria.

Deficiencies:

- The permit requires that the sample collection and flow readings should be conducted at the same time for loading calculations purposes. The plant was collecting the 24-hour flow reading with an eight hours lag (8:00am) from the time of collection of the 24-hour composite sample (4:00pm).
- An EPA instantaneous calibration check for the flow meter at outfall 001 indicated that the meter was recording 25 percent less than the actual flow, thus exceeding the ±10 percent threshold.

Regulatory Requirements: 40 CFR, Part 122.41, (e), Proper Operation and Maintenance: The permittee shall at all times properly operate and maintain all facilities and systems of treatment and control and related appurtenances which are installed or used by the permittee to achieve compliance with the conditions of this permit (40 CFR, Part 122.41, (e)). The permittee shall obtain accurate wastewater flow data to calculate mass loading (quantity) from measured concentrations of pollutants discharged as required by its NPDES permit. 40CFR part 122.41 (j) (1), Monitoring and Records states that samples and measurements taken for the purpose of monitoring shall be representative of the monitored activity. The difference between two stable flow totalizer readings (flow is steady for 10 minutes or more) should not exceed ±10 percent of the instantaneous flow measured at the primary device (NPDES Compliance Inspection Manual, EPA, 2004).

<u>Suggestion</u>: The Cornersville STP Operators should provide regular maintenance and cleaning of the effluent trough and check flow meter calibration on a daily basis.

3.4. Operations and Maintenance

The plant was operated and maintained by two certified operators, Mr. Kent Sweeton and Mrs. Denise Massey. The plant is staffed eight hours per day, five days per week and several hours on weekends. Maintenance activities were performed by the staff as needed. Laboratory equipment, such as the analytical balance, pH meter, and DO meter were observed in good condition. CBOD₅, E. coli, nutrients, TSS, and settleable solids analyses were performed at the Lewisburg Wastewater Treatment Plant laboratory.

Deficiencies:

- At the time of the inspection, the 1.2 MG equalization tank was near to exceeding capacity (Photo 4, page 29). The operator stated that the previous week the tank overflowed.
- The UV lamps did not appear to be clean.

Regulatory Requirement: 40 CFR, Part 122.41, (e), Proper Operation and Maintenance. The permittee shall at all times properly operate and maintain all facilities and systems of treatment and control and related appurtenances which are installed or used by the permittee to achieve compliance with the conditions of this permit (40 CFR, Part 122.41, (e)).

<u>Suggestion:</u> The city of Cornersville should evaluate the city sewer system for infiltration problems. A maintenance schedule is recommended to reduce the build up of grit and other materials in the bottom of the equalization tank. Ultimately, preliminary treatment processes including screening and grit removal should be installed at the head of the plant.

3.5. Sludge Disposal

The plant transports approximately 3,500 gallons of waste sludge every week in two 1,800 gallon truck tanks to the Lewisburg wastewater treatment plant. There, the sludge was treated in an aerobic digester tank, dewatered by a filter belt press and then disposed in a landfill.

4. SAMPLING

The permittee collected the required samples according to the sampling frequencies and sample types described in the NPDES permit. Influent flow proportional composite samples were collected for analyses of CBOD₅, TSS, total nitrogen, and total phosphorus using an ISCO 4700 automatic composite sampler. The influent sample was collected after the equalization tank.

Effluent flow proportional composite samples were collected with an ISCO 4700 automatic sampler and the tubing was installed at the discharge channel. The sample was set to collect approximately 200 ml sample aliquots. Samples for Outfall 001 were analyzed for total suspended solids (TSS), settleable solids, ammonia as N, total phosphorus, and carbonaceous biochemical oxygen demand (CBOD₅), at various frequencies. The permit monitoring requirements for ammonia varied depending of the season of the year (summer and winter).

Effluent grab samples were collected for pH, and dissolved oxygen (DO). A grab sample for *E. coli* was collected after the UV contact chamber. Samples were collected in accordance with the requirements specified in the NPDES Permit. The following table summarizes the NPDES permit sampling scheme for outfall 001:

Table 5: Permit Self-Monitoring Requirements

Parameters	Unit	Type	Frequency
Flow MGD		Continuous	Daily
CBOD5	mg/l	24-hr Composite	Three per Week
TSS	mg/l	Composite	Three per week
E. coli (Fecal)	Count/100ml	Grab	Three per Week
Total Nitrogen (N)	mg/l	Composite	Twice per Month
Ammonia (as N)	mg/l	Composite	Three per Week
T. Phosphorus	mg/l	Composite	Monthly
DO	mg/l	Grab	Five per Week
Settleable solids	ml/l	Composite	Five per Week
pН	SU	Grab	Five per Week

Deficiency:

The pH 7 buffer solution exceeded the expiration date (January, 2013).

Regulatory requirement: Calibration of continuous monitoring pH meters should be carried out at least daily against fresh buffers (minimum of two points) that bracket the expected sample pH and are approximately three pH units apart. (40 CFR, 136, Standard Methods, Method 4500-H⁺ B, 20th Edition, and EPA Methods, Method 150.1).

Deficiencies:

- The influent composite sampler tubing was clogged (Photo 5, page 30).
- The influent composite sampler was located after the equalization tank. Influent samples were collected after the equalization basin and do not reflect raw influent versus final effluent percent removals.

Regulatory Requirement: 40CFR part 122.41 (j) (1), Monitoring and Records: Samples and measurements taken for the purpose of monitoring shall be representative of the monitored activity. The influent automatic sampler should be properly maintained. The intake line should be checked regularly for rags and other debris that can cause clogging problems. Or, the line should be moved or repositioned in a location where rags and debris are not a problem. The influent automatic sampler should be moved to the raw influent pump station to collect a more representative sample of the raw influent coming into the plant.

4.1. Evaluation of Influent Data Results

The pollutant loadings and major processes were assessed. The strength of the influent wastewater for BOD5, nitrate-nitrite, total phosphorous, and Total Kjeldahl Nitrogen (TKN) was lower than the typical values. Table 6 shows a comparison of USEPA influent analytical results and typical influent wastewater concentrations. It's important to note that the sampling event was conducted when the plant was operated in storm mode, with a daily average discharge of 0.185 MGD. The facility's average flow is approximately 0.08 MGD.

Table 6: Comparison of Influent Results with Typical Values

D. D		Typical Value	*	City of Cornersville Influent
Parameter	Low	Medium	High	EPA Result
BOD ₅ (mg/L)	110	190	350	40 A
TSS (mg/L)	120	210	400	20
TKN (mg/L)	20	41	70	14
Ammonia (mg/L)	12	25	45	14
Nitrate +nitrite (mg/L)	0	0	0	0.30
Total Phosphorus (mg/L)	4	7	12	1.0

^{*}Wastewater Engineering: Treatment and Reuse 4th edition, Metcalf& Eddy, 2003. McGraw-Hill Companies, Inc. NYC³

4.1.1. USEPA Samples Results

The sampling event was conducted in accordance with the EPA Quality Assurance Project Plan for the City of Cornersville Sewage Treatment Plant, May 5, 2013. Sampling was conducted when the plant was running in storm mode because of raining events that took place in the previous week. Samples of the influent were collected at the influent 4" pipe (Photo 6, page 30). The influent sample was collected from the plant automatic sampler (ISCO 4700) as a split sample. Solids samples were collected in the sequencing batch reactor tank during the react (aeration) cycle. The *E. coli* and composite effluent samples were collected after the ultraviolet violet disinfection system. Figure 2 shows the locations of the wastewater samples collected during this evaluation. Influent and Effluent composite samples were split between EPA and the permittee, and a side by side comparison of facility and EPA analytical results was performed (Table 7). The results indicated satisfactory agreement between laboratories. See Attachment 5, SESD Final Analytical Results for a complete listing of analytical data. At the time of the

inspection the daily average effluent flow was 0.185 mgd. The NPDES permit of the plant was approved for a treatment facility with a design capacity of 0.1 mgd, thus exceeding the permit by 85 percent.

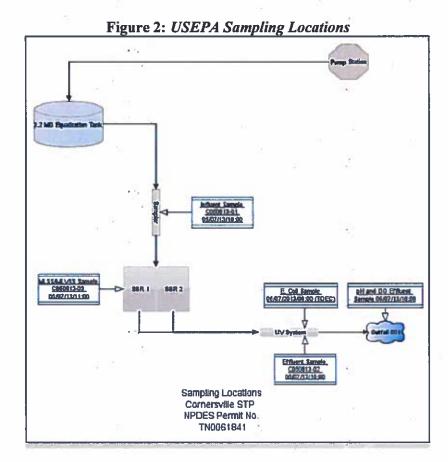


Table 7: USEPA and POTW Sampling Results (24 hour Composite)

Parameter	EPA Inf, mg/l	POTW Inf. mg/l	EPA Eff. mg/l	POTW Eff. mg/l	EPA Eff. Ib/d	POTW Eff. lb/d**	Removal Efficiency
CBOD ₅	40	30.9	5.3	3.33	8.2	4.72	87%
TSS	20	22	8.1	8.0	12.5	11.34	60%
Ammonia, as N	14	12.93	0.37	0.1	0.57	0.14	97%
TKN, mg/l	14	M. Street	1.4		2.2	-	90%
Nitrate +nitrite, mg/l	0.30		9.1	-	14.0		-
Total P, mg/l	2.0	A make	1.7		2.6		-
Settleable Solids, mL/I			0.50	0.1	W-		None
E. Coli, mpn/100ml	•		4***	3			None
DO, mg/l			10.7	11.0	Training I		
pH, S.U.			7.21	7.64		DE ELLES	None

Note: *USEPA Flow= 0.185 MGD was used to calculate the effluent loading (typical storm mode flow)

^{**}POTW Flow=0.170 MGD was used to calculate the effluent pollutant loading

^{***}E. Coli sample was analyzed by the TDEC Nashville Laboratory.

4.2. USEPA Sampling Methodology

All dissolved oxygen measurements were taken using YSI [®]550A Dissolved oxygen meter. pH was measured using an Orion pH meter. Flow measurement was checked at the effluent channel. All EPA sampling methods, measurements, and calibration were conducted in accordance with the following USEPA, Region 4, SESD procedures:

- Field Measurement of Dissolved Oxygen (SESDPROC-106-R2)
- Field pH Measurement (SEDPROC-100-R3)
- Wastewater Flow Measurement (SEDPROC-109-R3)
- Waste Water Sampling (SESDPROC-306-R3)
- In-situ Water Quality Monitoring (SESDPROC111-R3)
- Global Positioning System (SESDPROC-110-R3)
- Field Equipment Cleaning and Decontamination (SESDPROC-205-R2)

All EPA analyses, except *E. coli*, were conducted in accordance with the <u>Analytical Support Branch Laboratory Operations and Quality Assurance Manual</u>. February 15, 2013. The *E. coli* analysis was conducted by the Tennessee Department of Environmental Conservation (TDEC) Laboratory.

5. FACILITY DESIGN, OPERATION, AND PROCESS CONTROL ANALYSIS

The following sections described the evaluation of the unit processes performance and operation and discussion of the process control testing and operating parameters.

5.1. Preliminary Treatment

The SBR process is usually preceded by some type of preliminary treatment such as screening, communition or grit removal. The Cornersville STP does not have a preliminary treatment system. The plant operator stated that most of the influent comes from residential septic tanks that provide some sort of preliminary settling. However, commercial facilities that discharge their sanitary sewer did not provide any pre-treatment. The absence of a preliminary treatment may generate an accumulation of grit in the SBR tanks and create excessive wear of mechanical equipment due to the highly abrasive nature of grit and other materials.

5.2. Sequencing Batch Reactor Design and Operation

Each sequencing batch reactor has a volume of 4,719 ft³ and a depth of 13 ft in each tank (Photos 1 and 2, page 28). The SBRs process cycles are operated by a programmable logic controller (PLC). The facility's total cycle time is approximately 4 hours. Table 8 shows the Cornersville STP total SBRs cycle times and the total cycle time for an SBR with CBOD, suspended solids removal and nitrification.

Table 8: SBRs Cycle Times

SBR	Fill Mixed/ React (min)	Settle (min)	Decant (min)	Idle (min)	Total Cycle Time (Hrs)
Cornersville STP Normal Mode	131	63	66	1	4.35
Cornersville STP Storm Mode	85	19	. 64	. 1	2.81
SBR with Nitrification, BOD, TSS (recommended)*	180	60	60 (may vary)	1 (Variable, determined by flow rate)	5

^{*}SBRs for Nitrification and Nutrient Removal. U.S.EPA 832- R-92-002, September, 1992.

The permittee should evaluate increasing the total cycle time (fill, react, settle, and idle) to five hours and assess the ammonia-nitrogen removal efficiency of the system. For nitrification, the fill/react time cycle should be last three hours, including a react time of at least one hour (CSU, 2008). The following evaluations were conducted:

- Process control testing was performed to assess the quality of the mixed liquor in the biological process of the activated sludge. The following tests were performed for this evaluation: settlometer test, dissolved oxygen profile of the SBR tanks, microscopic examination and blanket depth measurement.
- Unit process operating parameters were also calculated to determine if the hydraulic and organic loadings were within the recommended range of operation and design for an SBR unit.

Table 9 includes a summary of the evaluation results.

Table 9: SBRs-Process Control Summary Results

Aeration Basin Parameters	EPA Results	POTW Results	POTW March Average	SBR Recommended Range	Comments
Aeration Volume, MG	0.07	0.07	0.07	SBR volume 1.2 to 2.0 times the daily flow*	Average flow is 0.08MGD
Number of SBR Cycles per day			5 cycles in SBR 1 and 6 cycles at SBR 2	2 to 6 cycles per day*	Normal
SSV ₃₀	350	-	-	· . •	Normal Settling
SSV ₃₀ Diluted	200	-	-	-	Rapid Settling, check MLSS
Hydraulic Detention Time, hrs	18	-	-	15-40**	Normal
Sludge Blanket depth, ft	7	-		Tanks are 13 ft depth each	High
MLSS, mg/l	4,300	-	4,937	2,000-5,000*	Within the higher range
Mixed Liquor Volatile Suspended Solids (MLVSS), mg/l	3,800	-		Greater than 80% of the total MLSS	88%

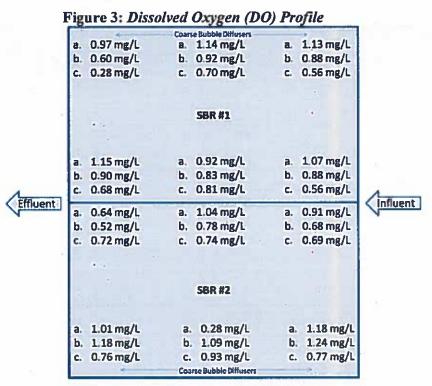
Aeration Basin Parameters	EPA Results	POTW Results	POTW March Average	SBR Recommended Range	Comments
DO in Aeration tank during React Cycle	0.28 to 1.15 mg/l	20		1.0-3.0 mg/l*	Low DO. For nitrification DO is 2.0 mg/l
Sludge Age, days	81	80	51	25 to 45*	Exceeded the range
F/M Ratio, lb BOD/day/lb MLVSS	0.03	0.02	0.02	0.04 to 0.10**	Low F/M ratio
Mean Cell Residence Time (MCRT), days	43	48	58	10-30**	high MCRT, high MLSS
Organic Loading, lbs/day-1,000ft ³	6.5	4.7	5.3	5-15**	-
Sludge Volume Index (SVI), ml/g	81			<100-150**	Good Settling
Mixed Liquor Color	Brown	Brown	-	Brown	"
Microscopic Exam	Amoeboids, Flagellates, and Stalked Ciliates		-	•	Normal sludge

Note: *Operation of Wastewater Treatment Plants Vol 1 and 2, 7th Edition, CSU, Sacramento, 2008.

**Wastewater Engineering: Treatment and Reuse 4th edition, Metcalf& Eddy, 2003.

The permittee used MLSS concentration of 3,500 mg/l as the target concentration to waste sludge from the SBRs. The EPA MLSS value of 4,300 mg/l exceeded the POTW MLSS target by 18.6 percent. In addition the average MLSS for the month of March (4,937 mg/l) exceeded the POTW target by 29 percent. MLSS concentrations higher than the target operating parameter may affect the TSS and settleable solids removal efficiency. The MLSS concentration was still within the recommended target range but it was high based on the plant sludge waste targets.

Figure 2 describes the D.O. Profile conducted at the facility. The study was performed during the react cycle of the SBRs. Measurement intervals were recorded at twelve locations, on the surface, at one foot and three feet depths. The DO profile indicated good mixing on the surface, but not adequate D.O. under the surface. The DO profile revealed low DO concentrations in the two SBR tanks, ranging from 0.28 to 1.15 mg/l. These DO concentrations, especially at 1 ft and 3 ft depths, create an unfavorable environment for the for the growth and removal action of the aerobic and nitrifying bacteria.



Note: a= Surface DO reading b= 1ft depth DO reading

c= 3ft depth DO reading

The sludge age was 71 days, which exceeds the recommended range of 25 to 45 days (CSU, 2008). The F/M ratio of 0.03 lbs BOD/day/lbs-MLVSS was below the design range of 0.04 to 0.10 lbs BOD/day/lbs-MLVSS. The plant should increase sludge waste to provide a healthy food to microorganism ratio.

The food to microorganism ratio was 0.02 lbs BOD/day-lbs MLVSS, a value considered low. High MLVSS concentrations affect the F/M ratio. The same occurred with the USEPA MCRT of 43 days and the POTW MCRT for the month of March of 58 days. The MCRT operating parameter for an SBR system is 10 to 30 days. For optimal CBOD₅ removal, DO concentrations in the aeration basins should be maintained between 1.0 to 3.0 mg/l (Metcalf and Eddy, 2003). The permittee needs to assess the amount in cubic feet of air applied to the SBRs per pound of BOD removed (ft³ air/BOD lbs removed). In addition, they need to determine the amount of air applied per gallon of wastewater treated. Typical air requirements parameters for a diffused aeration system range from 800 to 1500 ft³/lbs BOD-removed (EPA).

5.3. SBR Biological Nutrient Removal (BNR)

The permittee exceeded the limits of the NPDES permit for ammonia-nitrogen every month of 2012, except November, 2012 (Table 3). The USEPA result for ammonia-nitrogen concentration was 0.37 mg/l (Table 7, page 17); a value below the NPDES permit limit of 1.1 mg/l. Nitrate-nitrite concentration at the influent was 0.30 mg/l and 9.1 mg/l at the effluent. The conversion of ammonia to nitrate requires significant amounts of oxygen, 4.6 mg of O₂ per 1 mg of nitrogen oxidized.

Analytical results showed that nitrification occurred in the SBRs at the time of the inspection, given that ammonia was oxidized from 14 mg/l to 0.37 mg/l (Table 7, page 17), and converted to nitrate-nitrite. Historical data showed that the Cornersville STP influent ammonia-nitrogen concentrations were generally higher than during the EPA evaluation. For example, the average ammonia-nitrogen concentrations for the month of November and December 2012 were 29.4 mg/l and 19.8 mg/l, respectively.

Low DO concentrations affect the biological nutrient removal efficiency. Maximum nitrification rates occur at DO concentrations greater than 2 mg/1. Also Temperature, pH, and solids retention time (SRT) are important parameters in nitrification kinetics. The optimum temperature for nitrification is between 25 and 35°C.

Operators applied manually sodium bicarbonate to add alkalinity to the SBR biomass to control the pH. Nitrification results in the consumption of alkalinity. As alkalinity is consumed, pH decreases. The plant average pH in the activated sludge is 7. Optimum pH for nitrification is in the range of 7.5 to 9.0. Below pH 7 and above pH 9.8 the nitrification rate is less than 50 percent of the optimum (EPA, 1992).

On 2012, the permitteee contracted J.R. Wauford Company Consulting Engineers to evaluate the plant problems to comply with the NPDES permit limits. The consultant concluded that alkalinity in the Cornersville Wastewater Treatment Plant is adequate to allow nitrification to occur at its maximum rate and the reported dissolved oxygen concentrations in the reactors are rarely greater than 1.0 mg/1. The permittee provided a copy of the consultant report.

Low DO concentrations in the activated sludge and the results of the settlometer test revealed that gentrification was occurring. The sludge initially settled during the 30-minute settling test and then floated to the surface after 1.5 hours. Denitrification was likely occurring in the SBR tank (Photo 9, page 32). Some probable causes of sludge rising:

- The activated sludge process is being operated at a low F/M ratio. Low F/M ratio was confirmed
 by the results of the operating parameters evaluation.
- The sludge has been held for too long in the SBRs and consequently all the available dissolved oxygen has been used by the microorganisms. At the time of the inspection, the facility sludge age was 71 days and the MCRT was 43 days (See Table 6).

The denitrification process in the SBRSs may possibly affect the CBOD₅, TSS and settleable solids effluent concentrations, thus resulting in violations of the limits of the NPDES Permit.

5.4. UV Disinfection System Evaluation

A UV vertical lamp disinfection system was observed at the Cornersville STP. UV system was operational but the UV lamps did not appear to be clean (slime growth observed), and solids had accumulated in the channel. To disinfect the water, UV light must be intense enough to penetrate the cell walls of the pathogens. A dirty UV lamp has a reduced UV light density, thus affecting the efficiency of the UV system. UV disinfection with low-pressure lamps is not as effective for secondary effluent with TSS levels above 30 mg/l (USEPA, 1999). The permittee exceeded the

limits of the NPDES permit for *E. coli* numerous times in 2012 and February, 2013. Appropriate weekly maintenance of the UV system is required. Additionally, it is extremely critical that proper SBR operation and process controls be implemented to provide for adequate TSS and settleable solids removal.

6. EFFLUENT AND RECEIVING WATERS

The final effluent was clear. There were no visible solids, oil sheens or foam observed in the final effluent streams (Photo 11, page 33).

7. CONCLUSIONS

The Cornersville STP staff should focus on monitoring key operating parameters (MCRT, F/M, MLSS, sludge age) on a daily basis, interpret the data in relation to the effluent quality and sludge setteability, and conduct the necessary adjustments to meet the limits of the NPDES permit, specifically for CBOD₅, TSS, settleable solids, ammonia-nitrogen, and E. Coliform Bacteria. The solids inventory should be gradually reduced to obtain an MCRT that provides for adequate CBOD₅ removal and nitrification, as well as good settling characteristics. This will also bring the F/M ratio up, keeping the biological process more balanced and in an acceptable range of 0.4 to 0.10 lbs BOD/day-lbs MLVSS.

The SBRs diffuser system should be evaluated for performance and operation; including the blowers capacity to determine if the system has the capability to increase the air supply of the SBRs treatment processes. Higher DO levels are needed in the SBRs fill/react cycle (1 to 3 mg/l). The settlometer test showed a normal sludge setting but also showed that denitrification appears to be occurring at about 1.5 hours. To reduce the amount of gentrification that occurs during the decant cycle; the SBRs should be operated at the proper DO levels.

The Cornersville STP should evaluate the alternative of increasing the SBR cycle time to approximately five hours (from 4.35 hours) to provide the required time for proper nitrification. The fill/react cycle time should be at least 3 hours (CSU, 2008). The optimal decant time may vary depending on sludge settlability, compaction, and denitrification.

The permittee should consider installing an automatic lime or sodium bicarbonate feeders and pH probe in the SBRs units to control pH fluctuations. The average pH for the plant's activated sludge was 7. Optimum pH for nitrification is in the range of 7.5 to 9.0.

A cleaning and maintenance schedule for the UV system would improve the disinfection process and better help the plant meet the NPDES permit limits for *E.coli*. SBR operation and process controls should be adjusted to provide for adequate TSS removal of 30 mg/l or less (monthly average) and settleable solids removal to enhance the disinfection efficiency of the UV system.

The City of Cornersville should recalibrate the effluent secondary flow meter to achieve accurate readings. The discharge channel upstream of the flume needs to be cleaned and properly maintained to meet program requirements.

The permittee should monitor the average influent flow of the plant to identify if the plant exceeds the 85 percent of the design criteria on an average annual basis based on the previous twelve months of data. If the 85 percent of design criteria is exceeded, the permittee should start the planning and provide a schedule of improvements to begin within one year of the exceeding the plant design, to expand the plant's capacity.

The influent automatic sampler should be moved upstream of the equalization basin and should be properly maintained. The intake line should be checked regularly for rags and other debris that can cause clogging problems. Or, the line should be moved or repositioned in a location where rags and debris are not a problem.

The City should consider the installation of a preliminary treatment process, such as a bar screen and a grit removal system, to reduce the inorganic solids in the equalization tank and subsequent units. Screening and grit removal in the plant influent will help minimize solids throughout the plant. The equalization tank should be evaluated for a potential accumulation of solids that could be reducing its capacity. The City should also assess the city's sewer system to detect potential infiltration problems that appear to increase the influent flow of the plant during raining events.

Transcription errors on the DMRs should be corrected and can be prevented with the development of an in-house quality assurance/quality control process. DMRs should be reviewed at least 2 to 3 times to avoid future errors.

Additional training or continuing education for public works staff should be provided to improve the operation of the plant and help the plant get back in compliance.

8. REFERENCES

- 1. "Operation of Wastewater Treatment Plants" Volume 1 and 2, Seventh Edition, California State University (CSU), Sacramento, 2008
- Wastewater Technology Fact Sheet,"Sequencing Batch Reactors". Office of Water U.S.EPA 932-F-99-073, September, 1999.
- Wastewater Technology Fact Sheet,"Ultraviolet Disinfection". Office of Water U.S.EPA 832-F-99-064, September, 1999.
- 4. "Sequencing Batch Reactors for Nitrification and Nutrient Removal". Office of Water U.S.EPA 832-R-92-002, September, 1992.
- 5. "Biological Nutrients Removal Processes and Costs". Office of Water U.S.EPA-823-R-07-002, June, 2007.
- "Wastewater Engineering: Treatment and Reuse" 4th edition, Metcalf& Eddy, 2003.
 McGraw- Hill Companies, Inc. NY
- 7. "Activated Sludge Process Control Manual". USEPA SESD.
- 8. "Aerobic Biological Wastewater Treatment Facilities: Process Control Manual". Office of Water U.S.EPA III-A-524-77, March 1977.
- 9. "Field Manual for Performance Evaluation and Troubleshooting at Municipal Wastewater Treatment Facilities". Clean Water Consultants EPA contract-No. 68-01-4418, January, 1978.

Attachment 1: Photographic Log



Photo	taken	by:
Jairo	Castil	lo

Project Name: 13-0346-Cornersville STP

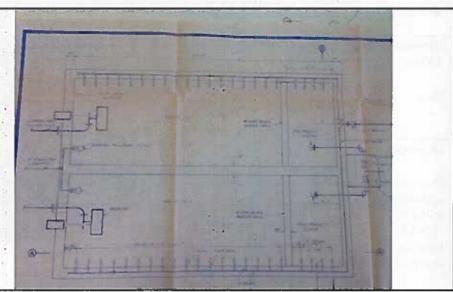
Photo No. Date: 5/7/2013

Direction Photo Taken:

South

Description:

Facility blueprint- two Sequencing Batch Reactors





PHOTOGRAPHIC LOG

Photo taken by: Jairo Castillo

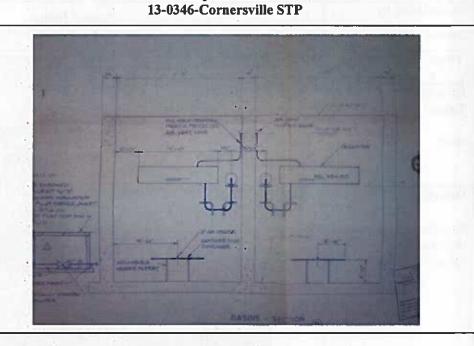
Photo No. Date: 5/7/2013

Direction Photo Taken:

South

Description:

Facility blueprint-Section view of the two sequencing atch reactors.



Project Name:



Photo taken by:

Jairo Castillo

Project Name: 13-0346-Cornersville STP

Photo No.

Date: 5/7/2013

Direction Photo Taken:

East

Description:

Pump station located approximately 450 feet to the north of the WWTP. Previous location of influent composite sampler.





PHOTOGRAPHIC LOG

Photo taken by:

Jairo Castillo

Photo No.

Date: 5/7/2013

Direction Photo Taken:

West

Description:

Equalization tank was observed near exceeding capacity.

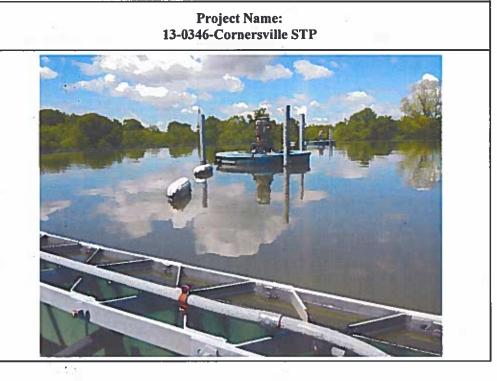




Photo taken by:

Jairo Castillo

Project Name: 13-0346-Cornersville STP

Photo No. 5

Date: 5/6/2013

Direction Photo Taken:

North

Description:

Influent sampler tubing clogged.





PHOTOGRAPHIC LOG

Photo taken by:

Jairo Castillo

Photo No.

Date: 5/6/2013

Direction Photo Taken:

North

Description:

Influent sampler tubing connected to the influent pipe before the inline electromagnetic flow meter.



Project Name:



Photo taken by:

Photo No.

Jairo Castillo

Date: 5/7/2013

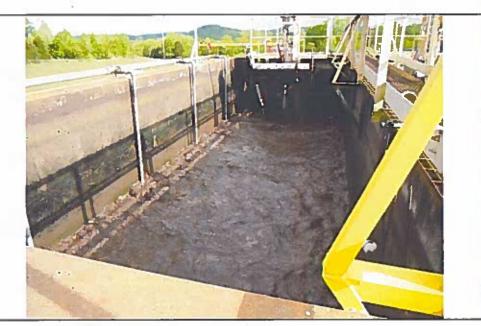
Direction Photo Taken:

South

Description:

SBR #2 during fill/mix cycle.

Project Name: 13-0346-Cornersville STP





PHOTOGRAPHIC LOG

Photo taken by: Jairo Castillo

Photo No.

Date: 5/7/2013

Direction Photo Taken:

South

Description:

SBR #1 during settling cycle



Project Name:



Photo taken by:

Jairo Castillo

Project Name: 13-0346-Cornersville STP

Photo No.

Date: 5/7/2013

Direction Photo Taken:

South

Description:

Settlometer test after 1.5 hours. Rising sludge is indicative of denitrification in the biomass.





PHOTOGRAPHIC LOG

Photo taken by:

Jairo Castillo

Photo No. Date: 10 5/7/2013

Direction Photo Taken:

East

Description:

Effluent automatic sampler (ISCO 4700).



Project Name:

13-0346-Cornersville STP



Photo taken by: Jairo Castillo Project Name: 13-0346-Cornersville STP

Photo No.

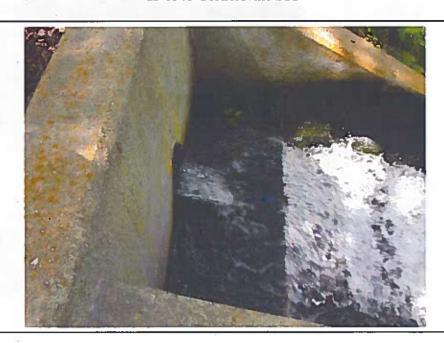
Date: 5/7/2013

Direction Photo Taken:

South

Description:

Cascade Aerator.
Effluent appeared clear.
No visible solids, oil sheens or foam observed in the final effluent streams.



						0.53			
		5							
		35		190					
		10 51 799							
						1000			
				62					
		1000							
		,							
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		. 18							
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. 4		19				3.5			
Attachm	ent 2: Prod	cess Conf	rol R	esults: S	Settlometer	Test a	nd Mier	oscopic F	valuation
				-54.000	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	= 1977		obtopie z	, , wilder
		4.7							
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		197							
						7,530			
	6					7.5			
			3	6		215.			
				- 54					
							89		
						539 8			
						W			
			25						

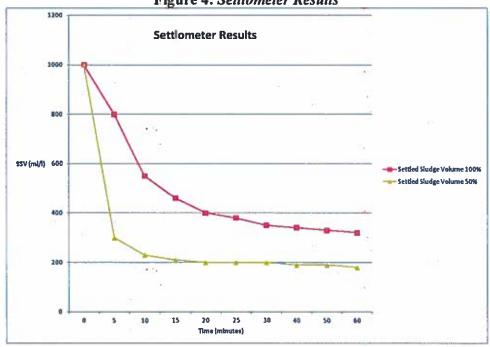
Settlometer Test

The settlometer test was performed to observe the settling characteristics of the mixed liquor, assess sludge quality and approximate age. A settlometer test was conducted at the plant using an undiluted 1,000 ml sample and a 50% diluted sample collected at the SBR during the react cycle (photo 9, page 31).

Table 10: Settlometer Test Results

Time (minutes)	Settled Sludge Volume (ml/l) 100 percent MLSS	Settled Sludge Volume (ml/l) 50 percent MLSS
0	1000	1000
5	800	300
10	550	230
15	460	210
20	400	200
25	380	200
30	350	200
40	340	190
50.	330	190
60	320	180

Figure 4: Settlometer Results



Microscopic Examination

Table 11: Microscopic Examination Results

Microorganisms	Quantity
Amoebae	10
Flagellates	o 10
Free swimming ciliates	4
Stalked ciliates	8
Rotifers	2
Worms	0
Nematodes	1

The predominance microorganisms were Amoeboids, Flagellates, and Stalked Ciliates. Free swimming ciliates were also observed in smaller quantities. Based on the microscopic examination, the sludge was normal.

Attachment 3: Operating Parameters Calculations

Cornersville STP Operating Parameters Calculations

USEPA calculated the operating parameters using three scenarios: USEPA diagnostic evaluation results, permittee's split sample results, and POTW March DMRs monthly average results for influent and Effluent CBOD₅ and TSS concentrations and MLSS Data for that month. March 2013 data was used to compare diagnostic evaluation data with the plant most recent data. MLSS and MLVSS concentrations were from SBR 1; calculations assumed that both tanks have the same concentrations (Historical data showed that there is a seven percent difference of MLSS concentrations between the SBRs). The following references were used for the calculations:

- "Operation of Wastewater Treatment Plants" Volume 1 and 2, Seventh Edition, California State University (CSU), Sacramento, 2008
- "Wastewater Engineering: Treatment and Reuse" 4th edition, Metcalf& Eddy, 2003. McGraw-Hill Companies, Inc. NY

EPA Data:

MLSS= 4,300 mg/l

BOD₅ influent= 40 mg/l

Influent Flow $(Q^0)=0.185 \text{ MGD}$

MLVSS plant average= 3,800 mg/l

Aeration basin volume= 0.070MG (2 tanks)

TSS_{inf}=20 mg/l

 $TSS_{eff} = 8.1 \text{ mg/l}$

POTW Data

V=0.070 MGD

CBOD₅ influent= 30.9 mg/l

Influent Flow (Q^0) = 0.170 MGD

Aeration basin volume= 0.070MG (2 tanks)

MLSS=4,300 mg/l (USEPA result used for calculations)

MLVSS=3800 mg/l (USEPA result used for calculations)

TSS_{inf}=22 mg/l

 $TSS_{eff} = 8.0 \text{ mg/l}$

POTW March average Data

CBOD₅ influent= 61.7 mg/l

Influent Flow (Q^0) = 0.098 MGD

Aeration basin volume= 0.070MG (2 tanks)

MLSS=4,937 mg/l

MLVSS=4,375 mg/l

 $TSS_{inf} = 69 \text{ mg/l}$

TSS_{cfl}= 4.5 mg/l

Hydraulic Retention Time for the SBR system (Only calculated for USEPA)

USEPA Data:

SVI=81 ml/g

Mass of solids at full volume=Mass of settled solids

 $VT*X=V_s*X_s$

V_t=Total volume, ft³

X=MLSS Concentration at full volume, mg/l

V_s=settled volume after decant, ft³

X_s= MLSS concentration in settled volume, mg/l

We need to solve the mass balance and determine the fill fraction/cycle.

a. Estimate X_s based on SVI value of 81 ml/g (measured onsite)

$$Xs = \frac{(10^3 \frac{mg}{g}) * (\frac{10^3 ml}{l})}{SVI, ml/g} = \frac{\left(10^3 \frac{mg}{g}\right) * (\frac{10^3 ml}{l})}{81 \frac{ml}{g}} = 12,346 \frac{mg}{l}$$

$$X = 4{,}300 \frac{\text{mg}}{\text{l}}$$
 (from sampling)

b. Settled Fraction

$$\frac{V_s}{V_T} = \frac{X}{X_s} = \frac{4,300 \frac{mg}{l}}{12,346 \frac{mg}{l}} = 0.35$$

Provide 20 percent liquid above the sludge blanket so that solids are not removed by decanting mechanism (Metcalf and Eddy, 2004).

$$\frac{V_S}{V_T} = 1.2 * (0.35) = 0.42$$

c. Calculate fill fraction

$$V_F + V_S = V_T$$

Where V_F=fill volume, ft³

$$\frac{V_F}{V_T} + \frac{V_S}{V_T} = 1.0$$

 $\frac{V_F}{V_T}$ = 1.0 - 0.42 = 0.58, using 0.5 as the fill fraction is acceptable.

d. Overall hydraulic detention time

Full liquid depth= 13ft

Decant depth= 0.5*(13ft)= 6.5 ft (Sludge judge measurements were 6-7 ft)

$$V_{T} = \frac{(4,719 \text{ ft}^{3})}{0.5} = 9,438 \frac{\text{ft}^{3}}{\text{tank}}$$

Overall Hydraulic Detention Time =
$$\frac{2 \tan ks * (9,438 \text{ ft}^3) * \left(24 \frac{\text{hr}}{\text{d}}\right) * 7.48 \text{ gal/ft}^3}{(185,000 \frac{\text{gal}}{\text{d}})} = 18 \text{ hours}$$

The hydraulic detention time of 18 days is within the recommended of 15 to 40 hours (Metcalf and Eddy, 2003).

Sludge Volumetric Index (USEPA only)

Data:

Settled Volume=350 ml/g at 30 min in the Settlometer test

$$SVI = \frac{\left(\text{Settled volume of sludge,} \frac{\text{ml}}{\text{L}} \text{ in 30 min}\right) \left(\frac{10^3 \text{mg}}{\text{g}}\right)}{\text{Suspended Solids,} \frac{\text{mg}}{\text{l}}}$$

$$SVI = \frac{\left(350 \frac{ml}{L}\right) \left(10^3 \frac{mg}{l}\right)}{4,300 \frac{mg}{l}} = 81 \frac{ml}{g}$$

A value of 100 ml/g is considered a good settling sludge, although SVI values below 100 are desired (Metcalf and Eddy, 2003).

Food-to-Microorganism Ratio (USEPA)

In equation form, the food-to-microorganism ratio is:

$$F/M_v = \frac{Q^0 S^0}{V X_v}$$

 F/M_v = food to microorganism ratio in volatile basis, lb BOD or COD per day of volatile suspended solids in aeration tank

 Q^0 = Influent wastewater stream flow rate (MGD)

S⁰= Influent wastewater BOD (or COD) concentration (mg/l)

V= aeration tank volume (MG)

X_V= volatile suspended solids concentration in aeration tank (mg/l)

Data

CBOD₅ influent= 40 mg/l

Influent Flow (Q^0) = 0.185 MGD

MLVSS plant average= 3,800 mg/l

Aeration basin volume= 0.070MG (2 tanks)

$$\frac{F}{M} = \frac{0.185 \text{ MGD} * 40 \frac{\text{mg}}{\text{l}} * 8.34 \frac{\text{lb}}{\text{gal}}}{0.070 \text{ MGD} * 3800 \frac{\text{mg}}{\text{l}} * 8.34 \frac{\text{lb}}{\text{gal}}} = 0.03 \text{ (low)}$$

A typical design parameter for an oxidation ditch activated sludge basin for the food-to-microorganism ratio (F/M) is 0.04-0.10 lb BOD/lb MLVSS-d (Metcalf and Eddy, 2003).

F/M Ratio (POTW)

Data

CBOD₅ influent= 30.9 mg/l

Influent Flow (Q⁰)= 0.170 MGD

MLVSS plant average= 3,800 mg/l

Aeration basin volume= 0.070MG (2 tanks)

$$\frac{F}{M} = \frac{0.172 \text{ MGD} * 30.9 \frac{\text{mg}}{\text{l}} * 8.34 \frac{\text{lb}}{\text{gal}}}{0.070 \text{ MGD} * 3800 \frac{\text{mg}}{\text{l}} * 8.34 \frac{\text{lb}}{\text{gal}}} = 0.02 \text{ (low)}$$

POTW F/M ratio (March 2013 average)

$$\frac{F}{M} = \frac{0.098 \text{ MGD} * 61.7 \frac{\text{mg}}{\text{l}} * 8.34 \frac{\text{lb}}{\text{gal}}}{0.070 \text{ MGD} * 4375 \frac{\text{mg}}{\text{l}} * 8.34 \frac{\text{lb}}{\text{gal}}} = 0.02 \text{ (low)}$$

Organic Loading Rate (USEPA)

$$OLR, \frac{lbs}{day - 1000ft^3} = \frac{Q_{in}, MGD * CBOD_5, \frac{mg}{l} * 8.34 * 1000}{Aeration Vol, ft^3}$$

$$OLR = \frac{0.185 MGD * 40 \frac{mg}{l} * 8.34 * 1000}{9,438 ft^{3}}$$

$$= 6.5 \frac{lbs}{day - 1,000ft^3}$$

An organic loading rate of 6.5 lbs/day-1,000 ft³ is within the recommended range of 5 to 15 lbs/day-1,000 ft³

Organic Loading Rate (POTW)

$$OLR, \frac{lbs}{dav - 1000ft^3} = \frac{Q_{in}, MGD * CBOD_5, \frac{mg}{l} * 8.34 * 1000}{Aeration Vol. ft^3}$$

$$OLR = \frac{0.172MGD * 30.9 \frac{mg}{l} * 8.34 * 1000}{9,438ft^{3}}$$

$$=4.7\frac{lbs}{day-1,000ft^3}$$

Organic Loading (POTW March 2013 average)

$$OLR, \frac{lbs}{day - 1000ft^3} = \frac{Q_{in}, MGD * CBOD_5, \frac{mg}{l} * 8.34 * 1000}{Aeration Vol, ft^3}$$

$$OLR = \frac{0.098MGD * 61.7 \frac{mg}{1} * 8.34 * 1000}{9.438ft^{3}}$$

$$=5.3\frac{lbs}{day-1,000ft^3}$$

Sludge Age (USEPA)

$$Sludge\ Age = \frac{Suspended\ Solids\ in\ Aerator,\ lbs}{Suspended\ Solids\ in\ Primary\ Effluent,\frac{lbs}{day}}$$

$$Sludge\ Age = \frac{MLSS, \frac{mg}{l}*Aerator\ Vol, MG*8.34\ lbs/gal}{Prim\ Eff, \frac{mg}{l}*Flow, MGD*8.34\frac{lbs}{gal}}$$

Sludge Age =
$$\frac{4,300\frac{mg_{*}}{l},0.070MG*8.34\frac{lbs}{gal}}{20\frac{mg}{l}*.185MGD*8.34\frac{lbs}{gal}} = 81 days (high)$$

The sludge age of 81 days exceeds the recommended range of 25 to 45 days (CSU, 2008).

Sludge Age (POTW)

Sludge Age =
$$\frac{\frac{4,300\frac{mg}{l}*0.070MG*8.34 \frac{lbs}{gal}}{22\frac{mg}{l}*0.172MGD*8.34\frac{lbs}{gal}} = 80 \text{ days (high)}$$

Sludge Age (POTW, March 2013 average)

Sludge Age =
$$\frac{4.937 \frac{mg}{l} * 0.070MG * 8.34 lbs/gal}{69 \frac{mg}{l} * 0.098MGD * 8.34 \frac{lbs}{gal}} = 51 days (high)$$

Mean Cell Residence Time (MCRT) (USEPA)

SBR Volume= 0.070MG

MLSS = 4,300 mg/l

Wasted Sludge Flow= 0.0005 MGD

Waste Sludge Suspended Solids Concentration= 10,694 mg/l

 $TSS_{eff} = 8.1 \text{ mg/l}$

$$MCRT = \frac{Suspended Solids (SS) in Aeration Systems, lbs}{SS Wasted, \frac{lbs}{day} + SS in Eff, lbs/day}$$

SS solids in Aerations Systems =
$$4,300 \frac{\text{mg}}{\text{l}} * 0.070 \text{ MG} * 8.34 \frac{\text{lbs}}{\text{day}} = 2,510 \text{ lbs}$$

SS Wasted,
$$\frac{\text{lbs}}{\text{day}} = 0.00052 \text{ MGD} * 10,694 \frac{\text{mg}}{\text{l}} * 8.34 \frac{\text{lbs}}{\text{gal}} = 46 \text{ lbs/day}$$

SS in Eff,
$$\frac{\text{lbs}}{\text{day}} = 0.184\text{MGD} * 8.1 \frac{\text{mg}}{\text{l}} * 8.34 = 12.42 \text{ lbs/day}$$

MCRT =
$$\frac{2,510 \text{ lbs}}{46 \frac{\text{lbs}}{\text{day}} + 12.4 \frac{\text{lbs}}{\text{day}}} = 43 \text{ days (high)}$$

The recommended design parameter for MCRT in oxidation ditch activated sludge is 15 to 30 days (Metcalf and Eddy, 2003).

MCRT (POTW)

$$MCRT = \frac{Suspended Solids (SS) in Aeration Systems, lbs}{SS Wasted, \frac{lbs}{day} + SS in Eff, lbs/day}$$

SS solids in Aerations Systems =
$$4,300 \frac{\text{mg}}{\text{l}} * 0.070 \text{ MG} * 8.34 \frac{\text{lbs}}{\text{day}} = 2510 \text{ lbs}$$

SS Wasted,
$$\frac{\text{lbs}}{\text{day}} = 0.00052 \text{ MGD} * 10,694 \frac{\text{mg}}{\text{l}} * 8.34 \frac{\text{lbs}}{\text{gal}} = 46 \text{ lbs/day}$$

SS in Eff,
$$\frac{\text{lbs}}{\text{day}} = 0.172 \text{MGD} * 4.5 \frac{\text{mg}}{\text{l}} * 8.34 = 6.45 \text{ lbs/day}$$

$$\text{MCRT} = \frac{2,510 \text{ lbs}}{46 \frac{\text{lbs}}{\text{day}} + 6.45 \frac{\text{lbs}}{\text{day}}} = 48 \text{ days (high)}$$

MCRT (POTW, March 2013 average)

$$MCRT = \frac{Suspended Solids (SS) in Aeration Systems, lbs}{SS Wasted, \frac{lbs}{day} + SS in Eff, lbs/day}$$

SS solids in Aerations Systems =
$$4,937 \frac{\text{mg}}{\text{l}} * 0.070 \text{ MG} * 8.34 \frac{\text{lbs}}{\text{day}} = 2,882 \text{ lbs}$$

SS Wasted,
$$\frac{\text{lbs}}{\text{day}} = 0.00052 \text{ MGD} * 10,694 \frac{\text{mg}}{\text{l}} * 8.34 \frac{\text{lbs}}{\text{gal}} = 46 \text{ lbs/day}$$

SS in Eff,
$$\frac{\text{lbs}}{\text{day}} = 0.098 \text{MGD} * 4.5 \frac{\text{mg}}{1} * 8.34 = 3.7 \text{ lbs/day}$$

$$\text{MCRT} = \frac{2.882 \text{ lbs}}{46 \frac{\text{lbs}}{\text{day}} + 3.7 \frac{\text{lbs}}{\text{day}}} = 58 \text{ days (high)}$$

Attachment 4: Cornersville STP, TDEC and SESD Laboratory Results

CORNERSVILLE WASTEWATER PLANT LAB TEST LOG# REPORT - 07-16-2013

CUSTOMER NAME : CORN - Tues DATE LOG NO. QTY 11.10 16.5 1 D. O. (Eff) 05-07-13 C13091 1 Temperature 1 S. S. (Eff) 0.1 7.64 1 pН E. Coli 3.1 BODS (Eff) BODS (Inf) Э.33 30.9 TSS (Eff) 8.0 1 TSS (Inf) 22.0 AMMONIA (Eff.) 0.1

LEWISBURG WATER & WASTEWATER LAB SAMPLE CHAIN OF CUSTODY

AMMONIA (Inf)

LOG NO. : C13091 SOURCE : CORN - Tues	DATE SAMPLED : 05-07-13
SAMPLE NAME : D. O. (Eff)	SAMPLE NAME : Temperature
SIGNATURE NO. : 1, 2	SIGNATURE NO. : 1, 2
SAMPLE NAME : S. S. (Eff)	SAMPLE NAME : pH
SIGNATURE NO. : \12	SIGNATURE NO. : 1, 2
SAMPLE NAME : E. Coli	SAMPLE NAME : BODS (Eff)
SIGNATURE NO. : \	SIGNATURE NO. :
SAMPLE NAME : BOD5 (Inf)	I SAMPLE NAME : TSS (Eff)
SIGNATURE NO. :)	! ! Signature No. : ハン
SAMPLE NAME : TSS (Inf)	I SAMPLE NAME : AMMONIA (Eff)
SIGNATURE NO. : 12	SIGNATURE NO. :
SAMPLE NAME : AMMONIA (Inf)	<u>.</u>
SIGNATURE NO. :\	
ANALYST AND SAMPLER SIGNATURES :	
1. SAMPLER Omaney	
2. ANALYST D. Marsey	
3. ANALYST CTOLO	
4 ANAT UST	



DIVISION OF LABORATORY SERVICES

Jackson Regional Laboratory 295 Summar Drive Jackson, TN 38301 731-426-0686

Shelby County Laboratory 814 Jefferson Avenue Memphis, TN 38105 901-544-7555

Sent To:

Ryan Owens TDEC-DWR

Columbia Field Office 1421 Hampshire Pike Columbia, TN 38401

Sampling Agency: TDEC: Division of Water Resources

J - Estimated value between MDL and MQL MDL - Method Detection Limit MQL - Method Quantitation Limit U - Undetected

Knoxville Regional Laboratory 2101 Medical Center Way Knoxville, TN 37920 865-549-5201

Nashville Central Laboratory 630 Hart Lane Nashville, TN 37243 615-262-6300

> Lab ID: N00010401 Nashville Central Laboratory



TDEC-DWR.N00010401.E

This is to certify that the following results were determined using good laboratory practices and in accordance with federal or state approved methodologies.

Analytical Supervisor

Page 1 of 2

Lab-Sample Number:

N00010401001

Field Determinations

pH:

Project Name: Sample Description:

CORNERSVILLE STP

Chlorine, residual: Conductivity:

Temperature:

Dissolved Oxygen:

Other:

Station No.:

Project Site No.:

NOT GIVEN

Flow:

CFS

Date/Time Collected: Sampler's Name:

Sampler Project Name: NPDES

05/07/2013 09:00 D. LOGSDON MARSHALL - 59

County: Sample Matrix:

Water

EFO: Sampling Agency: Columbia TDEC-DWR

Billing Code:

327.34-3082 TDEC-DWR Agency Invoiced:

TDEC-DWR

Send Report To:

Priority Date: Date/Time Received:

05/07/2013 12:15

Received By:

P Arjmandi

TEST: Escherichia Coli PERFORMING LAB: Nashville	ME	THOD:		
ANALYTE	RESULT	UNITS	ANALYZED BY:	DATE
Method Citation	SM 9223B		V Jordan	5/7/2013
E. Coli Result	4.0	MPN/100ml	V Jordan	5/7/2013

State of Tennessee - Environmental Laboratories PLEASE PRINT LEGIBLY



Inorganic Analysis

PROJECT/SITE NO.	PROJECT NAME	NPDEC	* Metals	
STATION NUMBER		MARSHAII 59	aluminum, Al	Laboratory Number 1000 104 01
DESCRIPTION COR	weasville STP		antimony, Sb	Branch Lab Number
STREAM MILE	DEPTH	MATRIX WATER		
COLLECTED: DATE		ME 9AM	arsenic, As	Chain of Custody and Supplemental Information
BAMPLER'S NAME(printed)		41L -7/4/M	barium, Ba	Only one chain of custody form is required per sample
	DEC/WAR /CEFO BILLIN	c cons 3=3.2 // 5 ==	beryllium, Be	set or point (if all collected at the sarpe time)
F PRIORITY, DATE NEEDED	DELTINA / CEPO BILLIN	G CODE 327.34-3082	cadmium, Cd	1, Collected by D. Lugs den
SEND REPORT TO:	9 . 0		calcium, Ca	Date 5-7-13 Time 9 Am
	RYAN OWENS		chromium, Cr	Delivered to STATE LAB
CONTACT HAZARD		(16:50 16:22 [] 성의 경임이 프로시아)	cobalt, Co	Dato 5-7-13 Time /2-43/p.m
* Env. Microbiology			copper, Cu	2. Received by
coliform, fecal*	" General inorganics	Gen. Inorganics (con't)	iron, Fe	Date Time
coliform, total*	acidity as CaCO ₃ °	oil and grease	lead, Pb	Delivered to
strep, fecal*	alkalinity as CaCO ₃ *	orthophosphate, total*	magnesium, Mg	Date Time
⟨ E. Coli*	alkalinity, phen. as CaCO ₃ *	oxygen, dissolved*	manganese, Mn	3. Received by
Enterococcus*	BOD, 5-day	pH	mercury, Hg	Date Time
Litter acoccus.	CBOD, 5-day	phenols, total	nickel, NI	Delivered to
* Appliant Decomp	boron	phosphate, total	potassium, K	Date / /Time
Amblent Parameters	chloride*	residue, dissolved*	selenium, Se	4. Received in Lab by Commount
COD.	chlorine, residual*	residue, settleable*	silver, Ag	Date 5/7/13 Time /2, 4
coliform, fecal	chromium, hexavalent	residue, suspended*	sodium, Na	Logged in by
conductivity*	COD*	residue, total*	thalium, TI	Date Time
hardness, total as CaCO3*	color, apparent*	silica*	vanadium, V	Tune
nitrogen, ammonia	color, true*	sulfate*	zinc, Zn	Additional Information
nitrogen, NO ₃ & NO ₂	conductivity*	sulfide, total*		1. Approximate volume of sample 250 mg/
nitrogen, total Kjeldahl	cyanide	Toc.		Zoo Ail
phosphate, total	flash point"	turbidity*		
pН	fluoride*	percent solids		2. Nearest town or city CORNEASVILLE
residue, dissolved*	hardness, Ca as CaCO ₃ *	* Ashestos	• TCLP	3 Others present at collection
residue, suspended"	hardness, total as CaCO ₃ °	bulk asbestos	arsenic, As	JAIRO CASTILO VALENZUALA BILL SALOS
arsenic, As	hydrocarbons, total	other microscopic	barium, Ba	4. Number of other samples collected at same time at
cadmium, Cd	MBAS*		cadmium, Cd	4 1 7 6
chromium, Cr	nitrogen, ammonia	* Other	chromium, Cr	this point None
copper, Cu	nitrogen, nitrate*		lead, Pb	
lead, Pb	nitrogen, nitrite*	43	mercury, Hg	5 Eight collection recognition by all the set
mercury, Hg	nitrogen, NO ₃ & NO ₂	[2] 1252 2	nickel, Ni	5. Field collection procedure, handling and/or preservation of this sample 57.7% SOP
nickel, Ni	nitrogen, total Kjeldahl		solenium, Se	preservation of this sample 5GTE SOP
zinc, Zn	nitrogen, total organic		silver, Ag	
ienoles analyses performed on	y on water		laurant v.A.	6. Mode of transportation to lab locked STATE
EIE D DETERMINE				
FIELD DETERMINATIONS	Temperature			vehrele
pH	Chlorino, rosidual	FOR	<01	7. Sample sealed by
Conductivay	Other	1-0R	21/4	8. Date sample scaled
Dissolved Oxygen				9. Remarks
				S. DOMENS



Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700 D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

June 7, 2013

4SESD-ASB

MEMORANDUM

SUBJECT:

FINAL Analytical Report

Project: 13-0346, Cornersville STP

Compliance Monitoring

FROM:

Roberta Howes

ICS Chemist

THRU:

Mike Wasko, Chief

ASB Inorganic Chemistry Section

TO:

Jairo Castillo

Attached are the final results for the analytical groups listed below. These analyses were performed in accordance with the Analytical Support Branch's (ASB) Laboratory Operations and Quality Assurance Manual (ASB LOQAM) found at www.epa.gov/region4/sesd/asbsop. Any unique project data quality objectives specified in writing by the data requestor have also been incorporated into the data unless otherwise noted in the Report Narrative. Chemistry data have been verified based on the ASB LOQAM specifications and have been qualified by this laboratory if the applicable quality control criteria were not met. Verification is defined in Section 5.2 of the ASB LOQAM. For a listing of specific data qualifiers and explanations, please refer to the Data Qualifier Definitions included in this report. The reported results are accurate within the limits of the method(s) and are representative only of the samples as received by the laboratory.

Analyses Included in this report:		Method Used:	Accreditations:
Classical/Nutrient Analyses (CNA)			
Ammonia/TKN		EPA 350.1 (Water)	ISO
Ammonia/TKN		EPA 351.2 (Water)	ISO
Demand	Y.	SM 5210B (Water)	ISO
Nitrate and/or Nitrite		EPA 353.2 (Water)	ISO
Phosphorous		EPA 365.1 (Water)	150
Solids		SM 2540E (Water)	None
Solids		SM 2540F (Water)	None
Solids		USGS 1-3765-85 (Water)	ISO

6/7/13 12:45



Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700 D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

Sample Disposal Policy

Because of the laboratory's limited space for long term sample storage, our policy is to dispose of samples on a periodic schedule. Please note that within 60 days of this memo, the original samples and all sample extracts and/or sample digestates will be disposed of in accordance with applicable regulations. The 60-day sample disposal policy does not apply to criminal samples which are held until the laboratory is notified by the criminal investigators that case development and litigation are complete.

These samples may be held in the laboratory's custody for a longer period of time if you have a special project need. If you wish for the laboratory to hold samples beyond the 60-day period, please contact our Sample Control Coordinator by e-mail at R4SampleCustody@epa.gov, and provide a reason for holding samples beyond 60 days



Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700 D.A.R.T. ld: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

SAMPLES INCLUDED IN THIS REPORT

Project: 13-0346, Cornersville STP

Sample 1D	Laboratory ID	Matrix	Date Collected	Date Received
C050813-02	E131903-01	Wastewater	5/7/13 16:00	5/8/13 14:41
C050813-01	E131903-02	Wastewater	5/7/13 16:00	5/8/13 14:41
C050813-03	E131903-03	Wastewater	5/7/13 11:00	5/8/13 14:41



Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700 D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

DATA QUALIFIER DEFINITIONS

- U The analyte was not detected at or above the reporting limit.
- A The analyte was analyzed in replicate. Reported value is an average value of the replicates.

ACRONYMS AND ABBREVIATIONS

- CAS Chemical Abstracts Service
 - Note: Analytes with no known CAS identifiers have been assigned codes beginning with "E", the EPA ID as assigned by the EPA Substance Registry System (www.epa.gov/srs), or beginning with "R4-", a unique identifier assigned by the EPA Region 4 Jaboratory.
- ISO The test, if analyzed after June 26, 2012, is accredited under the EPA Region 4 ASB's ISO/IEC 17025 accreditation issued by ANSI-ASQ National Accreditation Board/ACLASS. Refer to certificate and scope of accreditation AT-1691.
- MDL Method Detection Limit The minimum concentration of a substance (an analyte) that can be measured and reported with a 99% confidence that the analyte concentration is greater than zero.
- MRL Minimum Reporting Limit Analyte concentration that corresponds to the lowest demonstrated level of acceptable quantitation. The MRL is sample-specific and accounts for preparation weights and volumes, dilutions, and moisture content of soil/sediments.
- TIC Tentatively Identified Compound An analyte identified based on a match with the instrument software's mass spectral library. A calibration standard has not been analyzed to confirm the compound's identification or the estimated concentration reported.



Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700 D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

Classical/Nutrient Analyses

Project: 13-0346, Cornersville STP

Sample ID: <u>C050813-02</u>

Lab ID: <u>E131903-01</u>

Station ID: CEFF001

Matrix: Wastewater

Date Collected: 5/7/13 16:00

CAS Number	Analyte	Results Qualifiers	Unites	MRL	Frepared	Inalyzed	Method
7664-41-7	Ammonia as N	0.37	mg/L	0.050	\$/13/13 9:11	5/14/13 15:16	EPA 350.1
E17148461	Fotal Kjeldahl Nitrogen	1,4	mg/L	0.050	5/13/13 10,41	5/15/13 11:21	EPA 351.2
E1640614	BOD, 5 Day, Carbonaccous	5.3 A	mg/L	2.0	5/09/13 9:02	5/14/13 13.13	SM 5210B
E701177	Nitrate/Nitrite as N	9.1	mg/L	0.50	5/23/L3 11-05	5/23/13 11.05	EPA 353.2
7723-14-0	Total Phosphorus	1.7	mg/L	1.0	3/14/13 14.09	5/14/13 18:26	EPA 365.1
E1642842	Settleable Solids	0.50 U	mL/L	0.50	5/09/13 11:15	5/09/13 12:00	SM 2540F
E1642818	Total Suspended Solids	8.1	mg/L	4.0	5/14/13 14:40	5/14/13 14:40	USGS 1-3765-85



Region 4 Science and Ecosystem Support Division
980 College Station Road, Athens, Georgia 30605-2700
D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

Classical/Nutrient Analyses

Project: 13-0346, Cornersville STP

Sample ID: <u>C050813-01</u> Station ID: <u>CINF001</u> Lab ID: <u>E131903-02</u>

Matrix: Wastewater

Date Collected: 5/7/13 16:00

CAS Number	Analyte	Results Qualifiers	Units	MRL	Prepared	Analyzed	Meshod
7664-41-7	Aminonia as N	4 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 -	mg/L	0,50	\$/13/13 9:11	\$14/13 15/16	EPA 350.1
E17148461	Total Kjeldahl Nitrogen	14	mg/L	0.50	5/13/13 10:41	5/15/13 11:21	EPA 351.2
E1640614	BOD, 5 Day, Carbonaceous	40 A	mg/L	2.0	5/09/13 9:02	5/14/13 13:13	SM 5210R
5701177	Nitrate/Nitrite as N	0.30	mg/L	0.050	5/23/13 11.05	5/23/13 11:95	EPA 353.2
7723-14-0	Total Phosphorus	2.0	mg/L	1.0	5/14/13 14:09	5/14/13 18:26	BPA 365 1
E1642818	Total Suspended Solids	20	mg/L	4.0	5/14/13 14:40	5/14/13 14:40	USGS 1-3765-85



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Project: 13-0346, Cornersville STP - Reported by Roberta Howes

Classical/Nutrient Analyses

Project: 13-0346, Cornersville STP

Sample 1D: <u>C050813-03</u>

Station ID: CSBR

Lab ID: <u>E131903-03</u>

Matrix: Wastewater

Date Collected: 5/7/13 11:00

CAS Number	Analyte	Results Qualifiers	Units	MRL	Prepared	Inalyzed	Method
E1642818	Total Suspended Solids	4300	mg/L	4.0	5/L4/13 14:40	\$/[4/13 14:40	USGS 1-3765-85
E1640374	Volatile Suspended Solids	3800	mg/L	4.0	5/14/13 14:40	5/14/13 14.40	SM 2540E



Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700 D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

Classical/Nutrient Analyses (CNA) - Quality Control US-EPA, Region 4, SESD

Analyte	Result	Reporting Limit	Units	Spike Level	Source Result	%REC	#REC Limits	RPD	RPD Limit	Notes
Batch 1304080 - C 350.1 Ammonia					3.50					
Blank (1304080-BLK1)			•	Prepared: 0	04/19/13 At	nalvzed: 05	/14/13		8	
EPA 350.1				***************************************						
Ammonia as N	U	0.050	mg/L							
.CS (1304080-BS1)	4			Prepared: (H/19/13 Ai	ulyzed: 05	/14/13			
EPA 350.1						many - manufacture, super- par-	P (7-107) (E) (E) (E) (E) (E) (E) (E) (E) (E) (E			
Ammonia as N	0.93400	0.050	mg/L.	1,0000		93.4	90-110			
fatrix Spike (1304080-MS1)	Sour	ce: E131603-	01	Prepared: 0	4/19/13 Ar	nlyzed: 05	/14/13			
PA 350.1										
irimonia as N	2 0850	0.050	mg/l.	1.0000	1.1420	94.3	90-110		NJ.	
Intrix Spike Dup (1304080-MSD1)	Sour	:e: E131603-	01	Prepared: 0	14/19/13 An	alyzed: 05	/14/13			
PA 350.1										
mmonia os N	2.1270	0.050	mg/L	1.0000	1,1420	98.5	90-110	1.99	. 10	
IRL Verification (1304080-PS1)				Prepared: 0	4/19/13 An	alyzed: 05	/14/13	70		
PA 350.1									The Till that the Till the constraint the till a life to the till a li	irrianilariari de de como en
nimonia as N	0.052000	0.050	mg/L	0.050000		104	70-130			MRL
latch 1305040 - C SM5210 BOD				03						
lank (1305040-BLK1)				Prepared: 0	5/09/13 An	aluzeri: 05	ก่อกา		_:	_
M 5210B						,				
OD, 5 Day Carbonaceous	U	2.0	mg/L							
CS (1305040-BS1)				Prepared: 0	5/09/13 An	alyzed: 05/	(14/13			
M 5210B					arrife i eficiarrium francis amplemen a annum au	- I				
OD, 5 Day, Carbonaceous	177.50	20	mg/L	164.00		108	81-119			
uplicate (1305040-DUP1)	Source	e: E131903-	H	Prepared: 0.	5/09/13 An	alyzed: 05/	14/13			
M 5210B	**************************************					-		7-1-1-1- Au		
OD, 5 Day, Carbonaceous	4,4800	2.0	mg/L		5.3100			17.0	20	
atch 1305043 - C 2540 Solids										
CS (1305043-BS1)	*		÷	Prepared &	Analyzed: 0	5/09/13				
M 2540F		72822	7127257			ESSLO	12 14 12 14			
ettleable Solids	22 500	0.10	int/L	23.000		97.8	90-110			

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Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700 D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

Classical/Nutrient Analyses (CNA) - Quality Control US-EPA, Region 4, SESD

Result	Reporting Limit	Units	Spike Level	Source Result	%REC	**REC Limits	RPD	RPD Limit	Notes
			Prepared &	Analyzed	05/09/13	ramaras dadare mahrararadarara andrérna d	sses sens erus arrivolo-sul-t-t-t-t-t-t-t-t-t-t-t-t-t-t-t-t-t-t-t	dillerete statement and an annual statement of the statem	*
Son	rce: E131903-	01	Prepared &	Analyzed:	05/09/13		mbb		dans als als als sission als silvans all arthur
0,10000	0.10	mL/L		0.10000			0.00	10	
				(2)					
			Prepared: 0	5/13/13 At	nalyzed: 05	/15/13			a a ramaneous ries returnis less armelless dels a
U	0.050	ing/L							t
			Prepared; 0	5/13/13 Ai	nalyzed: 05	715/13			
d Philodolibelia discribitation de abrondos a monda brons monda de constante de la constante d									
2,4418	0.050	ing/L	2.3400		104	90-110			
Sou	ırce: E131903-	02	Prepared 0	15/13/13 A	nalyzed: 05	5/15/13			
17,773	0.50	mg/l.	1.0000	13.859	391	90-110			XM-
Sau	irce: E131903-	02	Prepared: 0	05/13/13 A	nalyzed: 0:	5/15/13			
18 014	0.50	tng/L	1 0000	13 859	-116	90-110	1,35	20	XM-
			Prepared: 0	05/13/13 A	nalyzed: 0	5/15/13	go- ay aga a agu a gu ga		
0.072200	0.050	mg/L	0,050000		144	70-1,30			MRL-2 QR-
			Premared &	k Analyzed	: 05/14/13				
en annes sels el destributes de relation del Mandado (1909-1949-1949)		and the standing of the standing of the							
U	0.010	mg/L							9
	2.4418 Sot 17.773 Sot 18.014	Source: E131903- 0.10000 0.10 U 0.050 2.4418 0.050 Source: E131903- 17.773 0.50 Source: E131903- 18.014 0.50 0.072200 0.050	Source: E131903-01 0.10000 0.10 mL/L 0.050 ing/L 2.4418 0.050 ing/L Source: E131903-02 17.773 0.50 mg/L Source: E131903-02 18.014 0.50 ing/L	Prepared & Source: E131903-01 Prepared & O.10000 O.10 mL/L	Prepared & Analyzed: Source: E131903-01	Prepared & Analyzed: 05/09/13 Source: E131903-01 Prepared & Analyzed: 05/09/13 0.10000 0.10 mL/L 0.10000 Prepared: 05/13/13 Analyzed: 05 U 0.050 mg/L Prepared: 05/13/13 Analyzed: 05 2.4418 0.050 mg/L 2.3400 104 Source: E131903-02 Prepared: 05/13/13 Analyzed: 05 17.773 0.50 mg/L 1.0000 13.859 391 Source: E131903-02 Prepared: 05/13/13 Analyzed: 05 18.014 0.50 mg/L 1.0000 13.859 416 Prepared: 05/13/13 Analyzed: 05 0.072200 0.050 mg/L 0.050000 144	Prepared & Analyzed: 05/09/13 Source: E131903-01	Prepared & Analyzed: 05/09/13 Source: E131903-01 Prepared & Analyzed: 05/09/13 0.10000 0.10 mL/L 0.10000 0.00 Prepared: 05/13/13 Analyzed: 05/15/13 U 0.050 mg/L Prepared: 05/13/13 Analyzed: 05/15/13 2.4418 0.050 mg/L 2.3400 103 90-110 Source: E131903-02 Prepared: 05/13/13 Analyzed: 05/15/13 17.773 0.50 mg/L 1.0000 13.859 391 90-110 Source: E131903-02 Prepared: 05/13/13 Analyzed: 05/15/13 18.014 0.50 mg/L 1.0000 13.859 416 90-110 135 Prepared: 05/13/13 Analyzed: 05/15/13 0.072200 0.050 mg/L 0.050000 144 70-130	Prepared & Analyzed: 05/09/13 Source: E131903-01 Prepared & Analyzed: 05/09/13 0.10000 0.10 mL/L 0.10000 0.00 10 Prepared: 05/13/13 Analyzed: 05/15/13 U 0.050 ing/L Prepared: 05/13/13 Analyzed: 05/15/13 2.4418 0.050 ing/L 2.3400 104 90-110 Source: E131903-02 Prepared: 05/13/13 Analyzed: 05/15/13 17.773 0.50 ing/L 1.0000 13.859 391 90-110 Source: E131903-02 Prepared: 05/13/13 Analyzed: 05/15/13 18.014 0.50 ing/L 1.0000 13.859 ill6 90-110 1.35 20 Prepared: 05/13/13 Analyzed: 05/15/13 0.072200 0.050 ing/L 0.050000 144 70-130



Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700 D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

Classical/Nutrient Analyses (CNA) - Quality Control US-EPA, Region 4, SESD

Amalyte	Result	Reponing Limit	Units	Spike Level	Source Result	**REC	%REC	nho.	RPD	21
naily te	Resuit	Limit	Onus	LCYEI	Kesuit	THREC	Limits	RPD	Limit	Notes
latch 1305060 - C 365.1 TPhos										
CS (1305060-BS1)				Prepared &	Analyzed	05/14/13				
EPA 365.1				100					1.+	
Total Phosphorus	0.37660	0.010	mg/L	0,40750		92.4	90-110			
latrix Spike (1305060-MS1)	Se	urce: E131903-	02	Prepared &	Analyzed:	05/14/13				
EPA 365.1										
Total Phosphorus	2.1700	1.0	mg/L	0.50000	1,9700	40.0	90-110			XVI
					2.5					
latrix Spike Dup (1305060-MSD1)	Sc	нигсе: Е131903-	02	Prepared &	Analyzed	05/14/13				
PA 365.1									30400	
ntal Phoephorus	2,3400	1.0	mg/L	0.50000	1.9700	74.0	90-110	7.54	10	XM
IRL Verification (1305060-PS1)				Prepared &	Analyzed:	05/14/13				
PA 365.1	The second secon					114 distantativ Sudet revists 400-1-31	9-1- His dasher-dashe en all transmitte arrange			
otal Phosphorus	0.0079000	0.010	mg/L	0.010000		79.0	70-130			MRL-2
latch 1305064 - C 2540 Solids										
lank (1305064-BLK1)				Prepared &	Analyzed:	05/14/13				
M 2540E										
'olatile Suspended Solids	u	4.0	mg/L							B-3, (
JSGS 1-3765-85										
otal Suspended Solids	U	4.0	12							1
• ** • • • • • • • • • • • • • • • • •	5657									
CS (1305064-BS1)				Prepared &	Analyzed: (05/14/13				
M 2540E						- Inmedia Individ				
olatife Suspended Solids	U	4.0	mg/L				90-110			NA, I
							100			
ISGS I-3765-85	102	,12		120000						
otal Suspended Solids	93.100	1.0	•	100.00		93.1	90-110			
CS Dup (1305064-BSD1)				Prepared &	Analyzed (05/14/13				
M 2540E				7.	,			-		
olatile Suspended Solids	U	40	mg/L				90-110		200	NA,
SGS I-3765-85										

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Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700

D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

Classical/Nutrient Analyses (CNA) - Quality Control US-EPA, Region 4, SESD

Sunday 1	Result	Reporting Limit	Units	Spike Level	Source Result	%REC	%REC Limits	RPD	RPD Limit	Notes
malyte	resuit	Littiit	Units	LEVEI	Result	AREC	Linns	KI-D	Lam	Muses
atch 1305064 - C 2540 Solids		7.099				-91				
Ouplicate (1305064-DUP1)	Sou	rce: E131903-	03	Prepared &	: Analyzed	05/14/13				
SM 2540E			5752							
Valatile Surperated Solids	3780.0	4 0	mg/l.		3800.0			0.528	10	
USGS 1-3765-85										
Total Suspended Solids	4412.0	4.0			4320.0			2.11	10	
MRL Verification (1305064-PS1)				Prepared &	Analyzed	05/14/13				
SM 2540E				1 toparea a	- Miniyasu	03/14/13				
Vulatite Suspended Solids	U	4.0	ing/L				70-130			NA, U
USGS 1-3765-85										
Total Suspended Solids	3.8000	4.0	-	5.0000		76.0	68-128			MRL-2
										,
Batch 1305102 - C 353.2 NO3-NO2										
Blank (1305102-BLK1)				Prepared &	Ł Analyzed	05/23/13				
EPA 353.2										
Nitrate/Nitrite as N	U	0.050	mg/L							,
Blank (1305102-BLK2)				Prepared &	k Analyzed	: 05/28/13				
EPA 353.2										
Nursie/Nitrite as N	υ	0.050	mg/L		•					,
Db1, /12/05103 DH 1/2\				Deannead	Pr A malesand	i nemula				
Blank (1305102-BLK3) EPA 353.2	- 10 - qui-fit d'en e stantade l'indressable se que é de flace sile-dominance de ma redicabet ve de	,		ricialeu c	& Analyzed	03129113				
Nitrate/Nitrite #4 N	U	0.050	mg/L							i
	×50	(34)		¥.						
I.CS (1305102-BS1)				Prepared &	& Analyzed	05/23/13				
EPA 353.2										
Nitrale/Nitrite os N	0.48980	0.050	mg/L	0.50000		98.0	90-110			
LCS (1305102-BS2)				Prepared A	& Analyzer	1: 05/28/13				
EPA 353.2										
Nitrate/Nitrite as N	0.51020	0.050	mg/L	0.50000		102	90-110			
1 CC (1102103 BC3)				Degrana 1	Ու գորժեսո	i nemnua				
LCS (1305102-BS3) EPA 353.2				rrepared	a Analyzet	1: 05/29/13		10 10 10		

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6/7/13 12:45



Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700 D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

Classical/Nutrient Analyses (CNA) - Quality Control US-EPA, Region 4, SESD

		Reporting		5pike	Source		%REC		RPD	
Analyte	Result	Limit	Units	Level	Result	*\$REC	Limits	RPD	Limit	Notes
Batch 1305102 - C 353.2 NO3-NO2										
.CS (1305102-BS3)		The state of the s	fr wa	Prepared &	& Analyzed:	05/29/13				13
Vitrate/Nitrite as N	0.49270	0.050	mg/L	0.50000		98.5	90-110			
Matrix Spike (1305102-MS1)	Sou	rce: E131808-	-09	Prepared &	k Analyzed:	05/23/13				V-114-0-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1
EPA 353.2										
Nitrate/Nitrite as N	0.72860	0,050	mg/L	0.50000	0.22490	101	90-110			
Matrix Spike (1305102-MS2)	Sou	rce: E131903-	02	Prepared &	k Analyzed:	05/23/13				
EPA 353.2										
Vitrate/Nitrite as N	0.78820	0.050	mg/l.	0.50000	0.29860	97.9	90-110			
Mateir Caller Dani (1205) 67 MCDM	e	F121808		N	آلسيا <u>س</u> ا	oemana				
Matrix Spike Dup (1305102-MSD1) EPA 353.2	30u	rce: F131808-		rrepared &	Analyzed:	U3/23/13				
EPA 333.2 Nitrate/Nitrite as N	0.74690	0.050	mg/L	0.50000	0.22490	104	90-110	2.48	10	
	4.74070	0.050	mg L	0.30000	0.22490	104	20-110	2.70		
Matrix Spike Dup (1305102-MSD2)	Sou	rce: E131903-	02	Prepared &	Analyzed:	05/23/13				
EPA 353.2										
Nitrate/Nitrite os N	0.81670	0.050	mg/L	0.50000	0.29860	104	90-110	3.55	10	
MRL Verification (1305102-PS1)				Denmarad &	Analyzed:	ns <i>r</i> 12/12				
EPA 353.2	Province of the second	tinks stadilish sharehida diselemberik sebih sheki she		i i c panca de	ritimy z.u.	(12/23/13	n affereir men ir imein ainma arth ar agus s			
Nitrate/Nitrite as N	0.039800	0.050	mg/L	0.050000		79.6	70-130			MRL-2.
			ÃĴ.			5				U
MR1. Verification (1305102-PS2)				Prenated &	: Analyzed:	05/28/13				
EPA 353.2		7		1 10 100 00	. renuly zeu.	03/20/13				
litrate/Nitrite as N	0.055200	0.050	mg/L	0.050000		110	70-130			MRL-2
			-				0.000			
IRL Verification (1305102-PS3)		4		Prepared &	: Analyzed:	05/29/13				
EPA 353.2										73.73.73.73.73.73.73.73.73.73.73.73.73.7
Nitrate/Nitrite as N	0.036100	0,050	mg/l.	0.050000		72 2	70-130			MRL-2,
										U



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UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

Region 4 Science and Ecosystem Support Division 980 College Station Road, Athens, Georgia 30605-2700 D.A.R.T. Id: 13-0346

Project: 13-0346, Cornersville STP - Reported by Roberta Howes

Notes and Definitions for QC Samples

U	The analyte was not detected at or above the reporting limit.	
B-3	Level in blank does not impact data quality	
MRL-2	MRL verification for Non-Potable Water matrix	
NA	Not Analyzed.	
QR-2	MRL verification recovery greater than upper control limits.	

Sample background/spike ratio higher than method evaluation criteria

Enclosure C

ENCLOSURE C

RIGHT TO ASSERT BUSINESS CONFIDENTIALITY CLAIMS (40 C.F.R. Part 2)

Except for effluent data, you may, if you desire, assert a business confidentiality claim as to any or all of the information that the EPA is requesting from you. The EPA regulation relating to business confidentiality claims is found at 40 C.F.R. Part 2.

If you assert such a claim for the requested information, the EPA will only disclose the information to the extent and under the procedures set out in the cited regulations. If no business confidentiality claim accompanies the information, the EPA may make the information available to the public without any further notice to you.

40 C.F.R. § 2.203(b). Method and time of asserting business confidentiality claim. A business which is submitting information to the EPA may assert a business confidentiality claim covering the information by placing on (or attaching to) the information, at the time it is submitted to the EPA, a cover sheet, stamped or typed legend, or other suitable form of notice employing language such as trade secret, proprietary, or company confidential. Allegedly confidential portions of otherwise non-confidential documents should be clearly identified by the business, and may be submitted separately to facilitate identification and handling by the EPA. If the business desires confidential treatment only until a certain date or until the occurrence of a certain event, the notice should so state.

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